

The background of the slide is a dense, abstract pattern of small, semi-transparent circles and triangles in various colors including red, green, blue, yellow, and purple. These shapes are scattered across the entire slide, creating a textured, mosaic-like effect.

Lecture 6

Inferring cell pseudotime from scRNAseq data

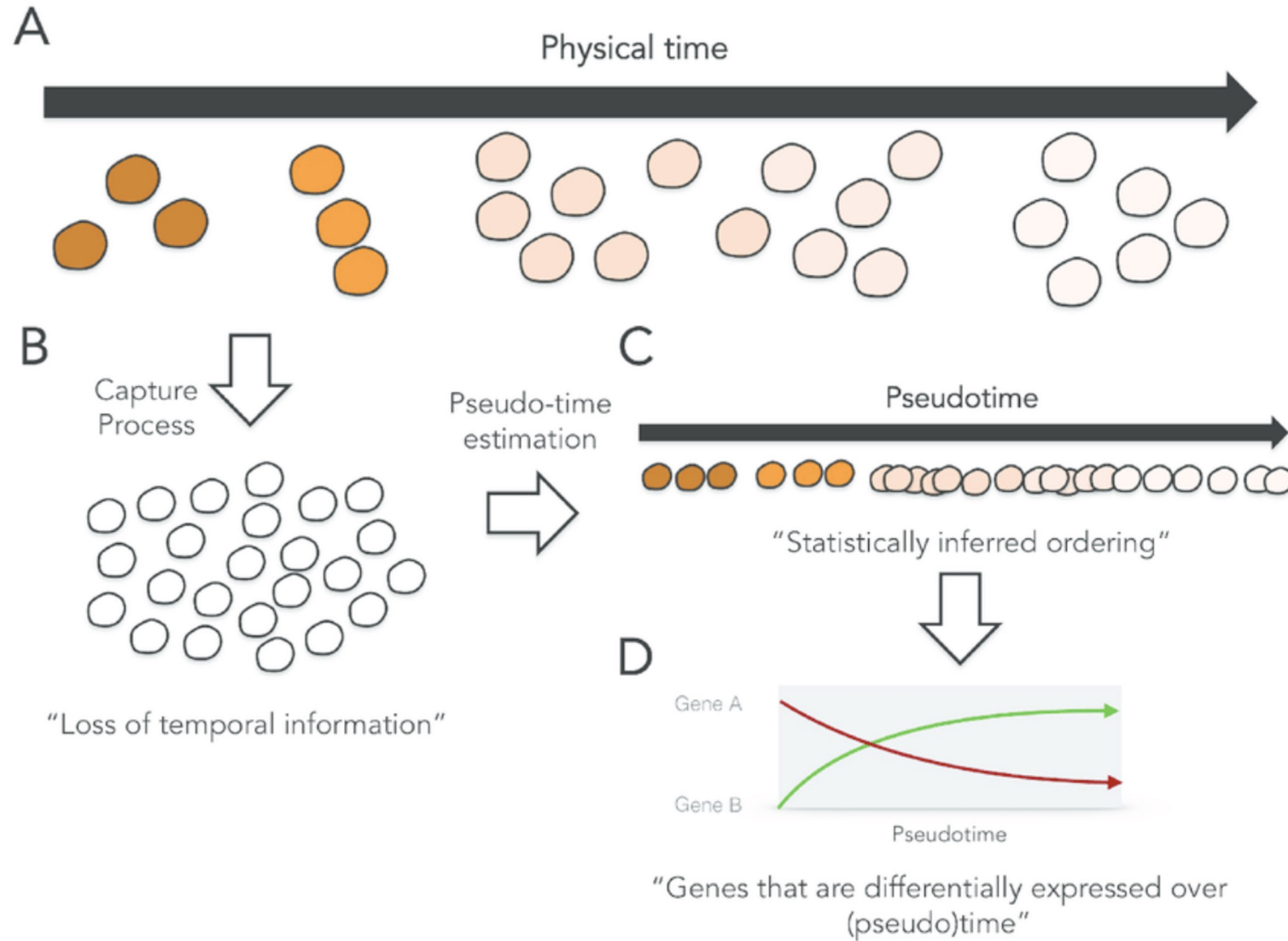
Physalia course 2023

—

Single-cell RNA-seq with R/Bioconductor

Instructors: Orr Ashenberg & Jacques Serizay

What is pseudotime?



What is pseudotime?

- Pseudotime is an abstract unit of progress: it's simply the distance between a cell and the start of the trajectory, measured along the shortest path
- The trajectory's total length is defined in terms of the total amount of transcriptional change that a cell undergoes as it moves from the starting state to the end state.
- **Pseudotime is a measure of how much progress an individual cell has made through a process (such as cell differentiation).**

What is pseudotime in single-cell RNA-seq?

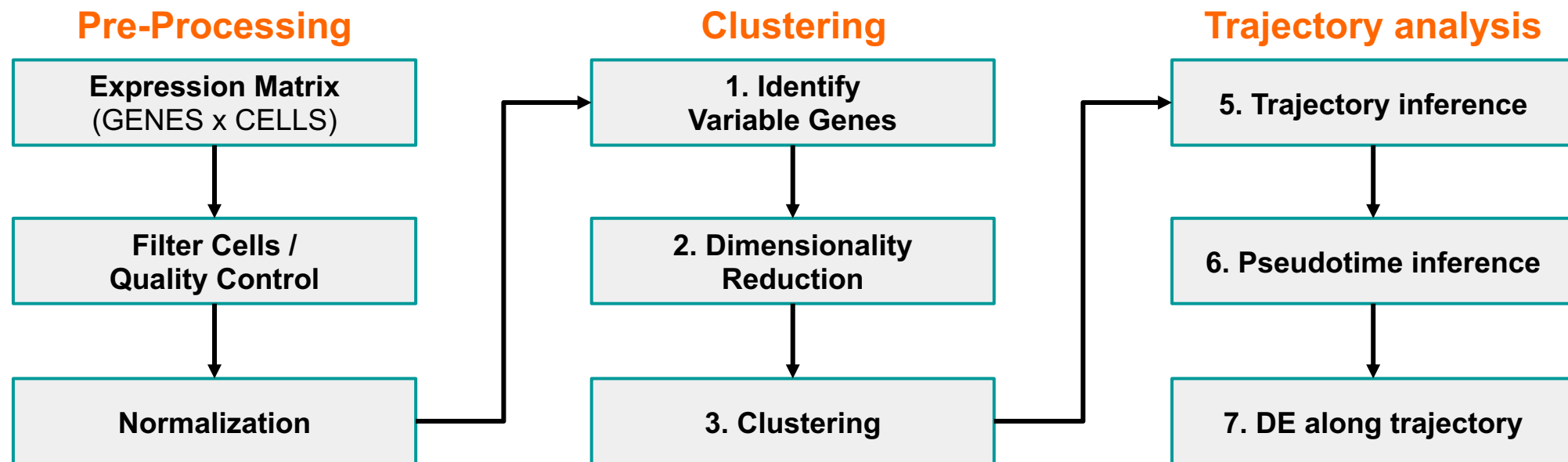
- In single-cell expression studies of processes such as cell differentiation, captured cells might be widely distributed in terms of progress.
- That is, in a population of cells captured at exactly the same time, some cells might be far along, while others might not yet even have begun the process.
- **By ordering each cell according to its progress along a learned trajectory, pseudotime inference alleviates the problems that arise due to asynchrony.**

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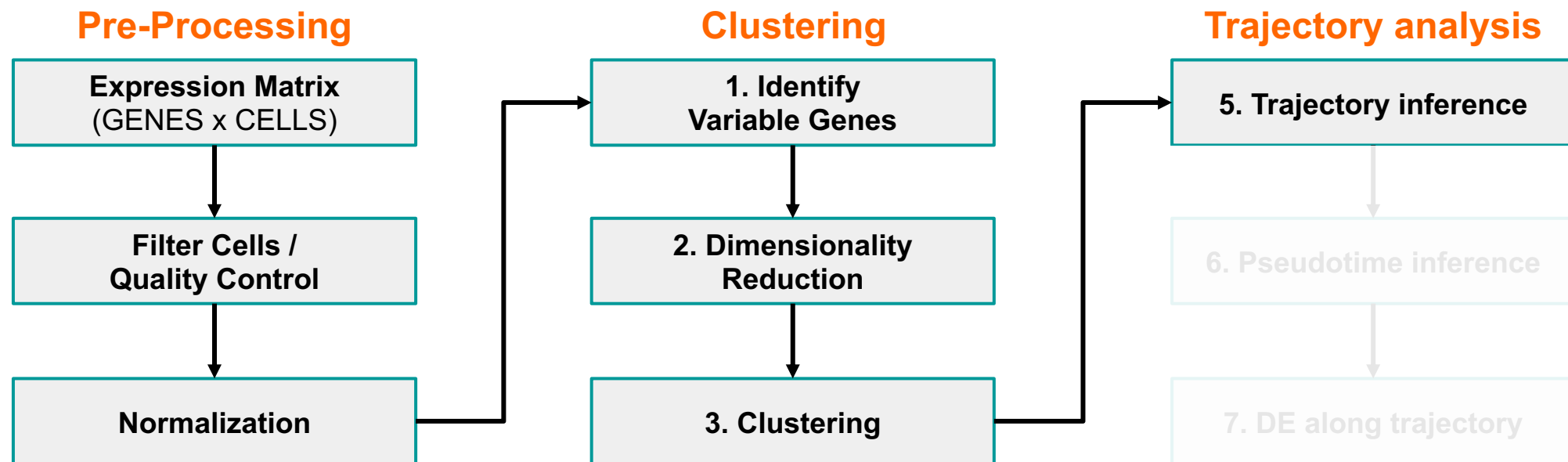
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This is why pseudotime and trajectory inference are largely overlapping.

Analysis workflow



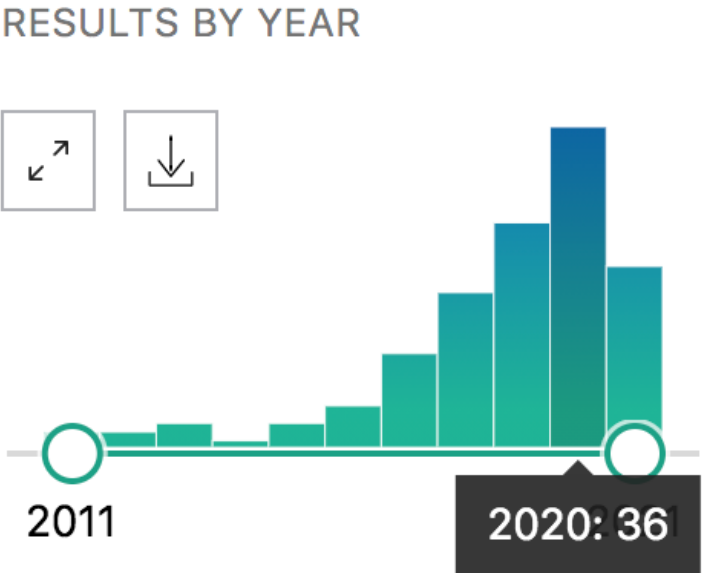
Analysis workflow



Trajectory inference (TI) tools

A new algorithm comes out ~ every other week...

Pubmed results for “**trajectory inference method single-cell**”:



Trajectory inference (TI) tools

A new algorithm comes out ~ every other week...

Pseudocell Tracer-A **method** for **inferring** dynamic **trajectories** using scRNAseq and its application to B cells undergoing immunoglobulin class switch recombination.

Reiman D, Manakkat Vijay GK, Xu H, Sonin A, Chen D, Salomonis N, Singh H, Khan AA.

PLoS Comput Biol. 2021 May 3;17(5):e1008094. doi: 10.1371/journal.pcbi.1008094. eCollection 2021 May.

PMID: 33939691 **Free PMC article.**

Single cell RNA sequencing (scRNAseq) can be used to **infer** a temporal ordering of cellular states. Current **methods** for the **inference** of cellular **trajectories** rely on unbiased dimensionality reduction techniques. ...

Slingshot: cell lineage and pseudotime inference for **single-cell** transcriptomics.

Street K, Risso D, Fletcher RB, Das D, Ngai J, Yosef N, Purdom E, Dudoit S.

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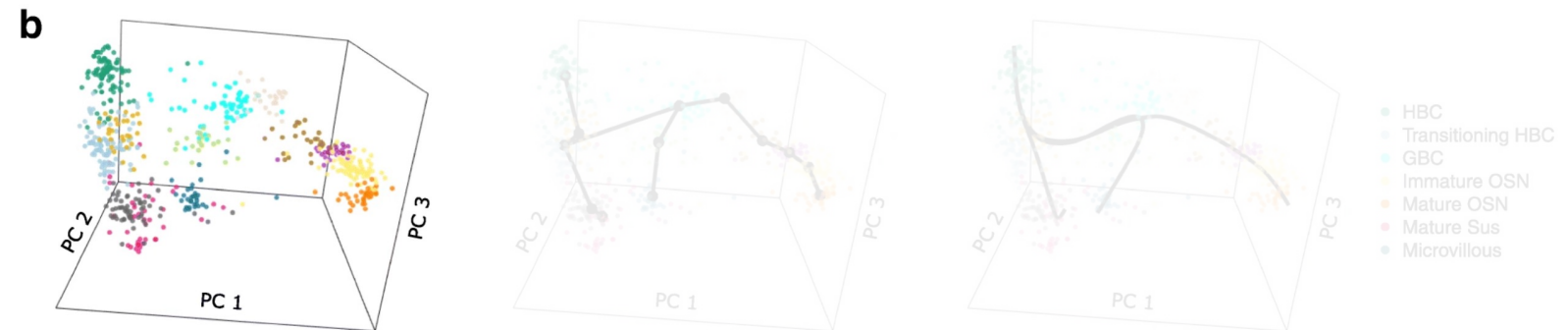
Stick to standards!!

QCed, troubleshooted, optimized and generic

Slingshot is one of the most widely used and robust approaches to infer trajectory in relatively simple datasets.

It works by:

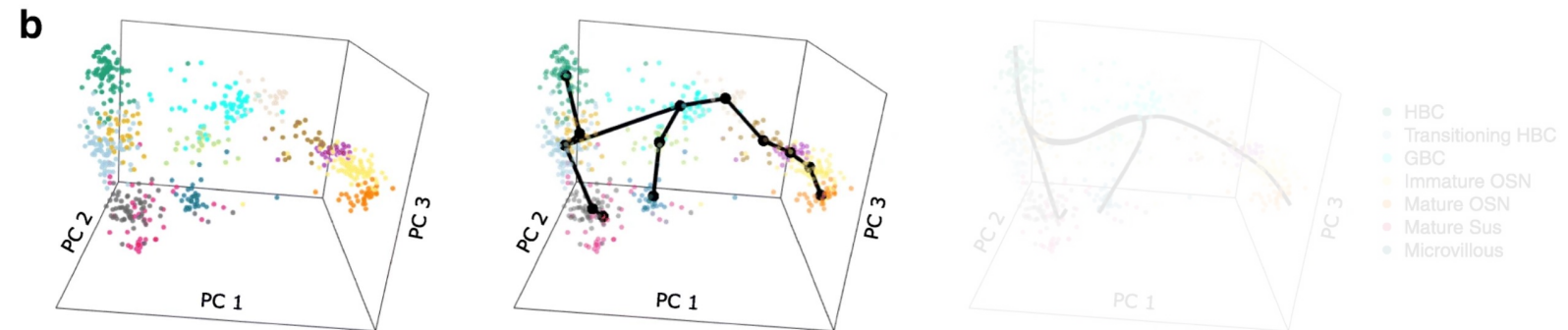
0. Get clustered data in a low-dimensional space



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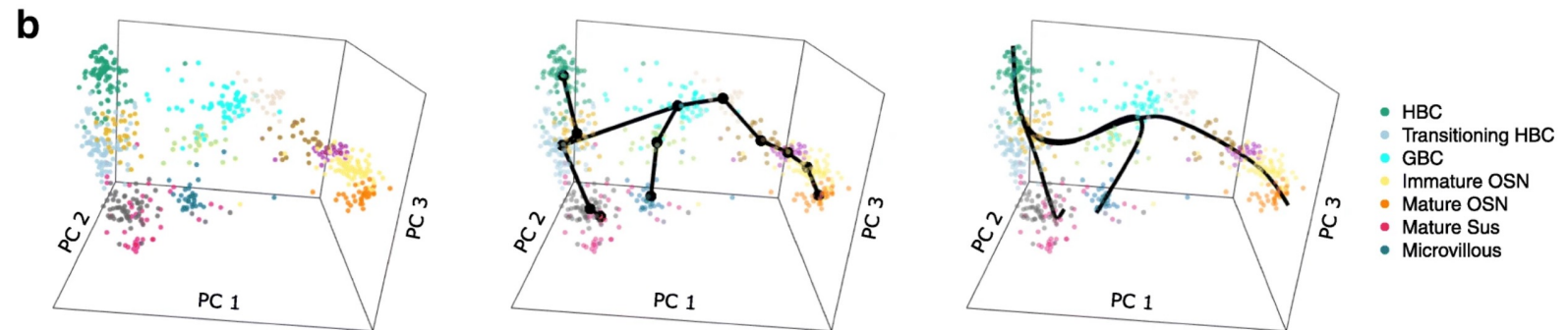
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1. Building a **minimum spanning tree** on the clusters



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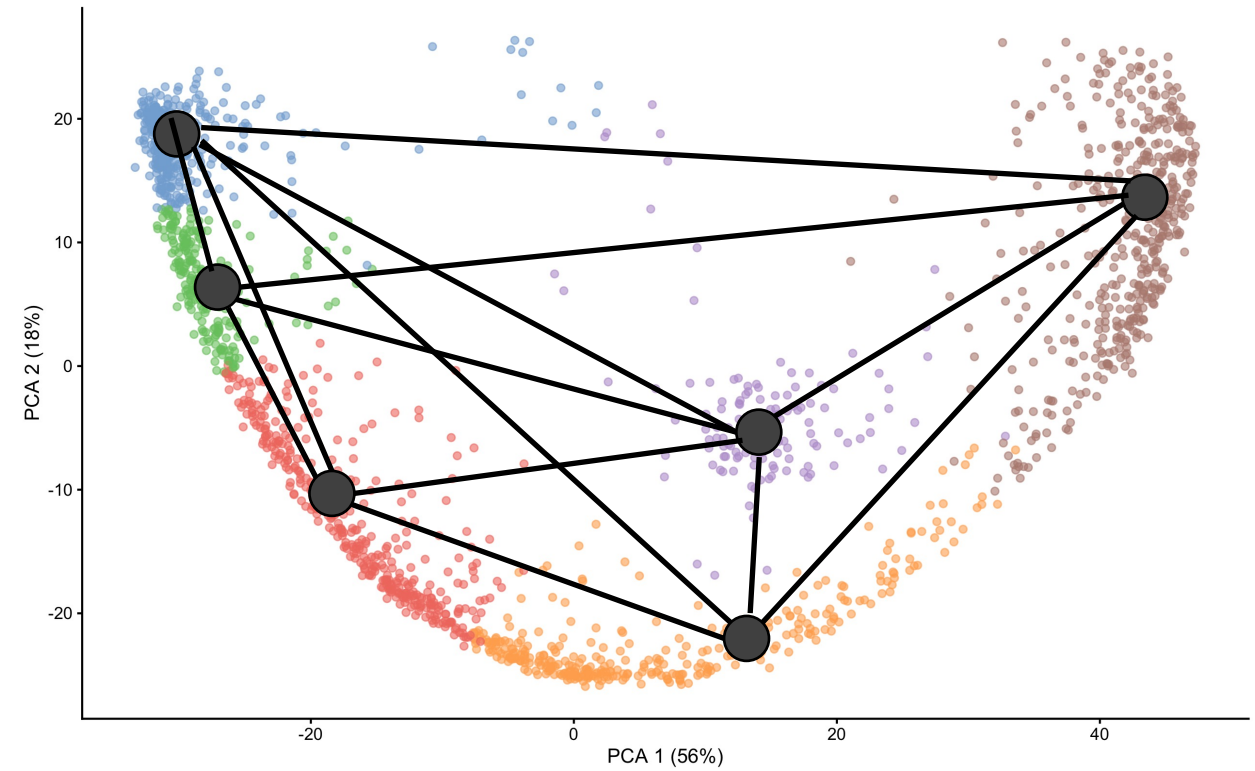
0. Get clustered data in a low-dimensional space
1. Building a **minimum spanning tree** on the clusters
2. Fit **principal curves** through the MST



Minimum spanning tree?!?

Or minimum weight spanning tree

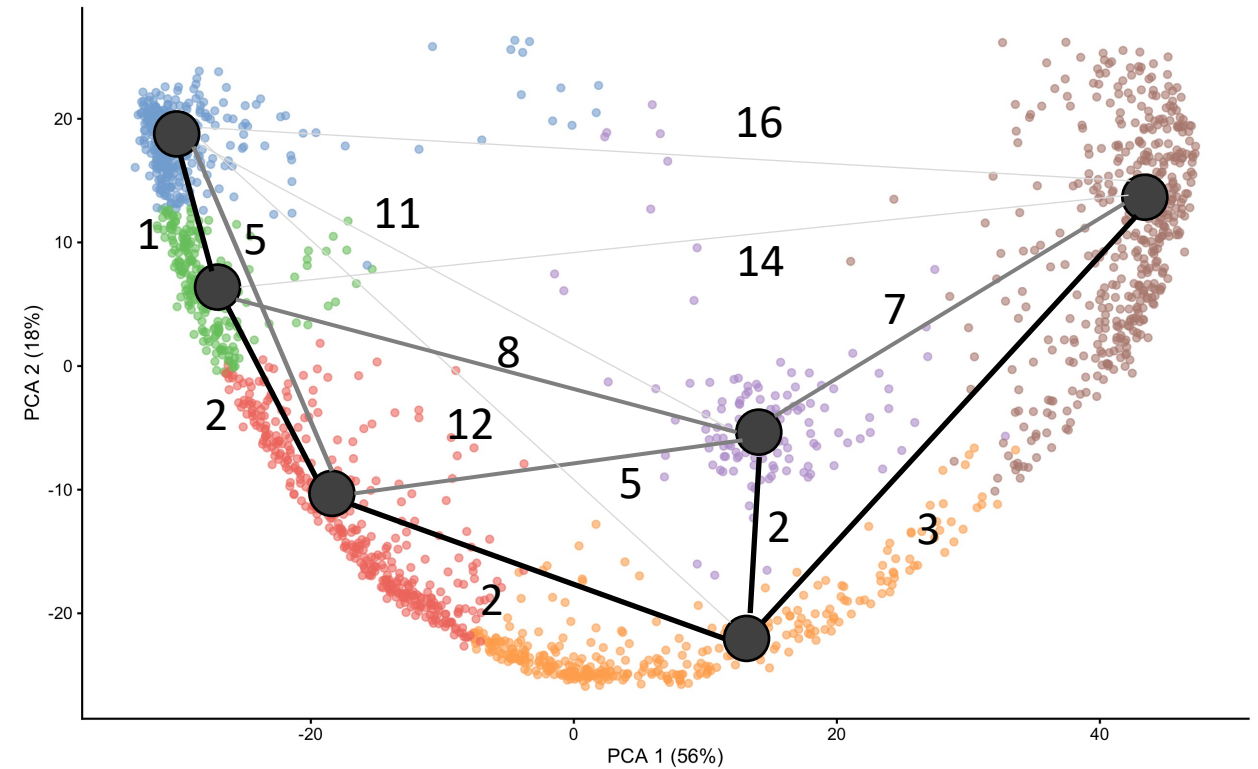
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Minimum spanning tree?!?

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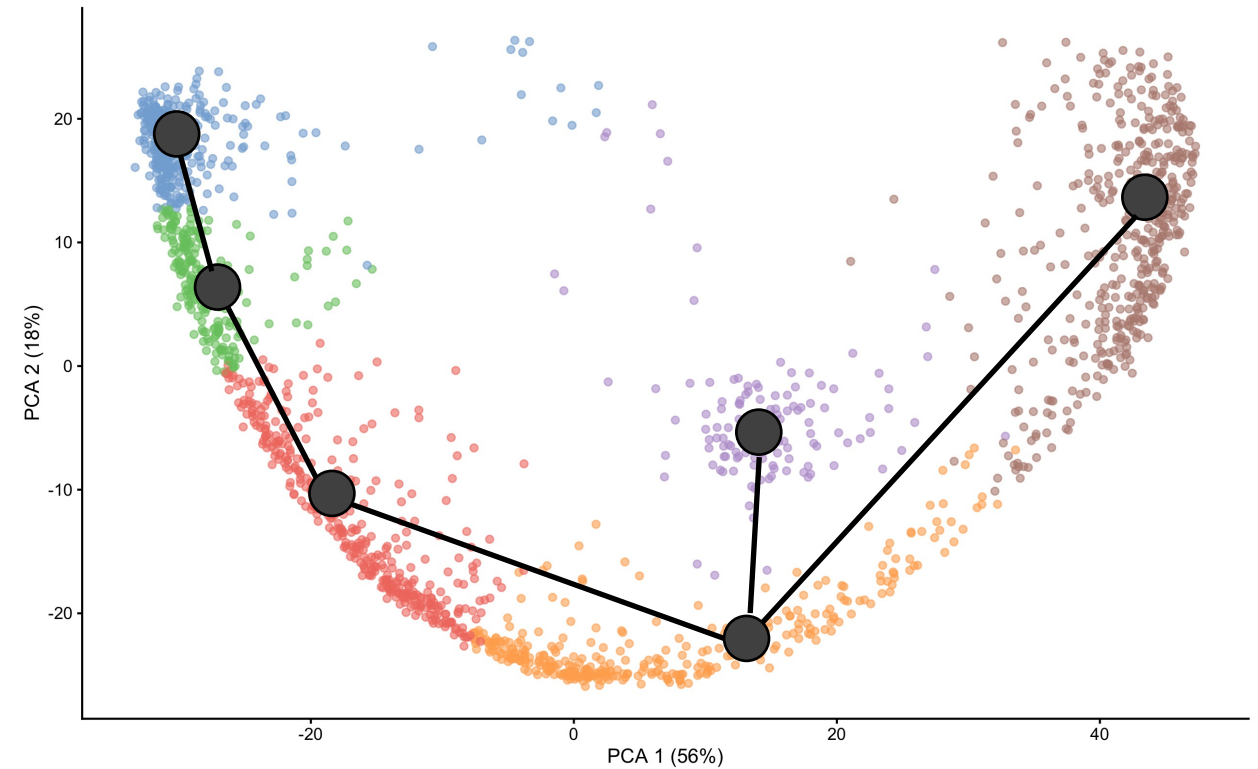
1. Build a graph with edges between each pair of clusters
2. Add weights to each edge according to proximity of the two clusters



Minimum spanning tree?!?

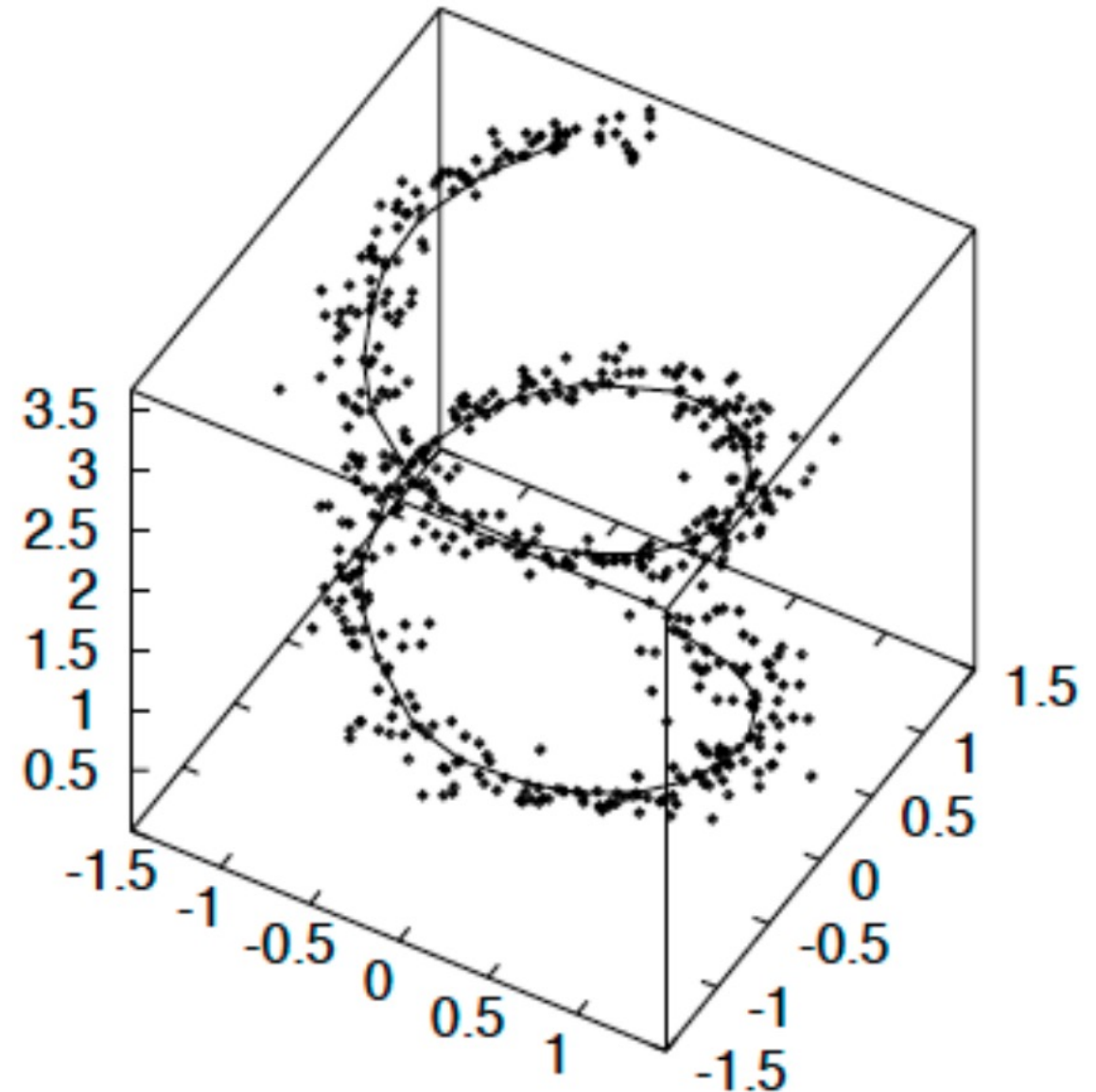
Or minimum weight spanning tree

1. Build a graph with edges between each pair of clusters
2. Add weights to each edge according to proximity of the two clusters
3. Find the shortest path between clusters



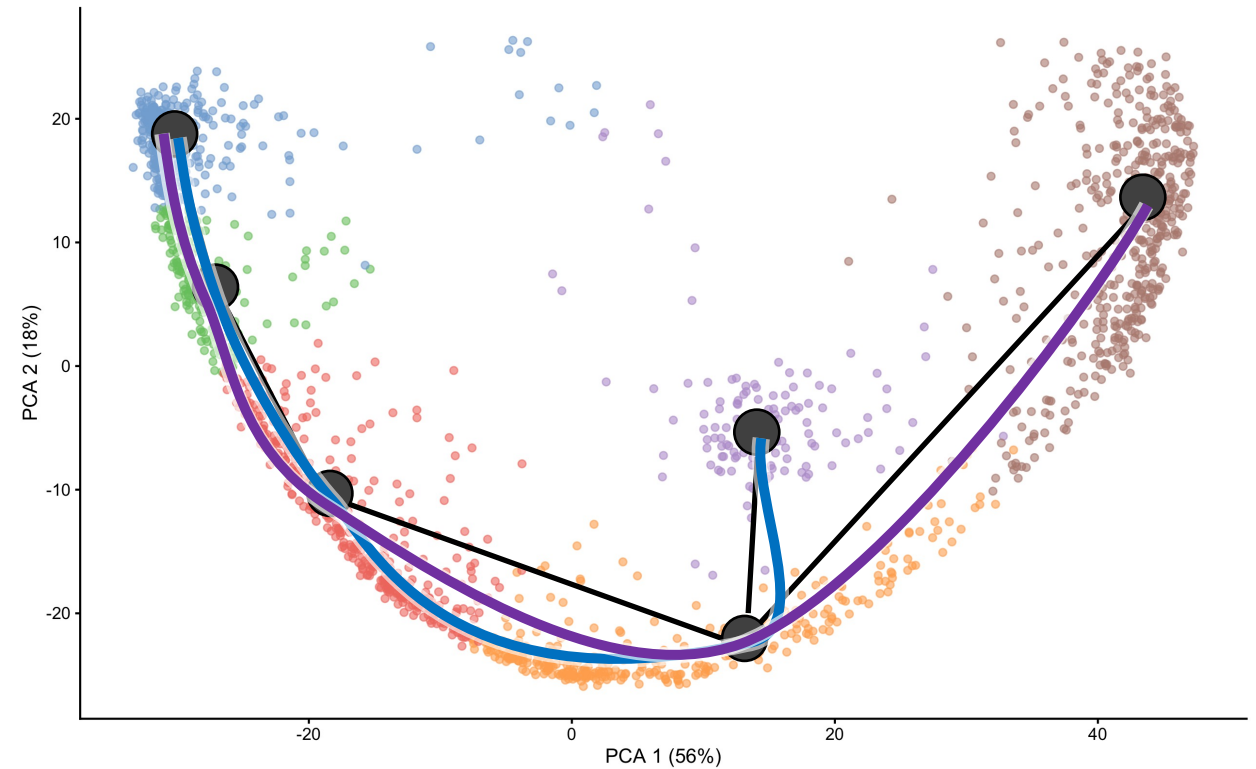
Principal curves?!?

A principal curve is a smooth, one-dimensional, curve that passes through the middle of a high-dimensional data set, providing a nonlinear summary of the data.



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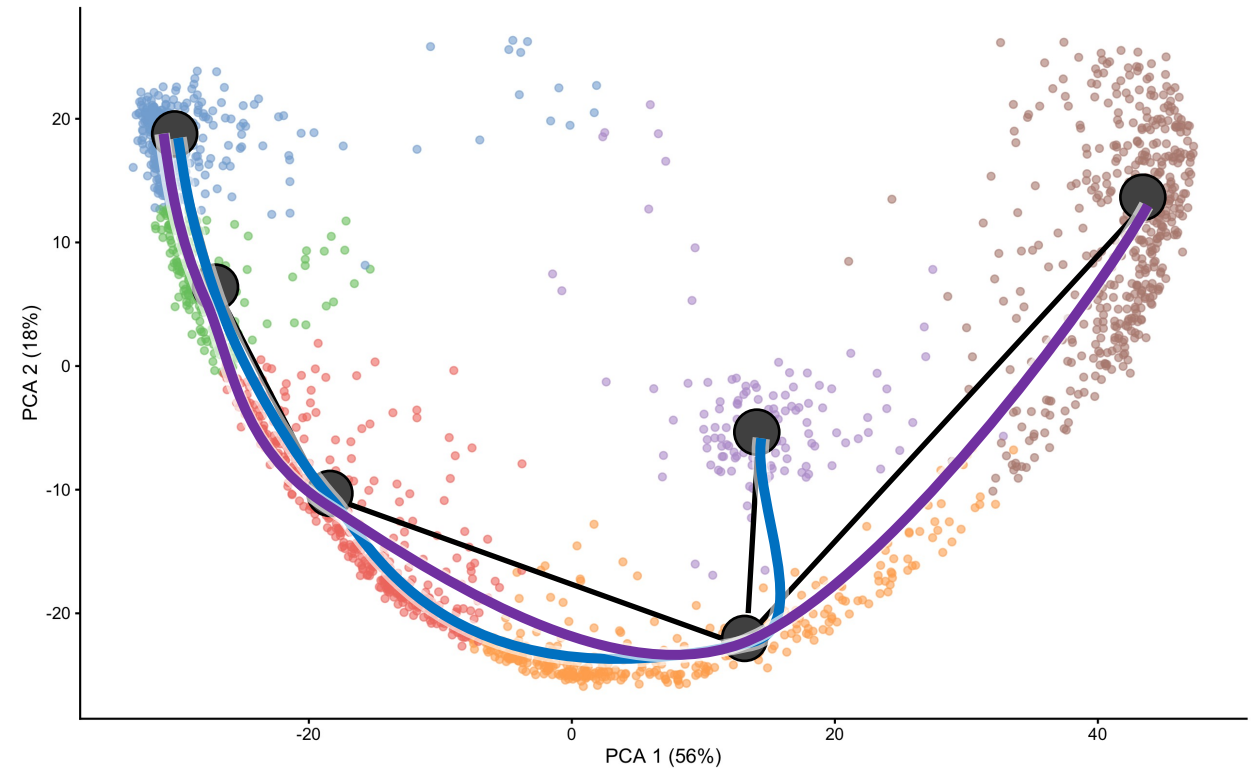


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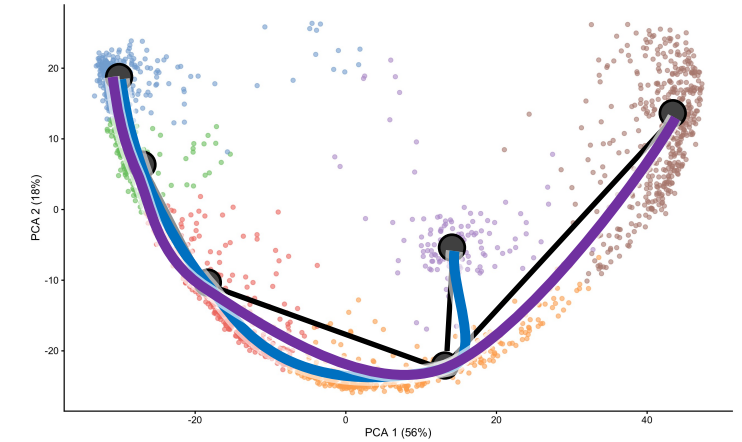
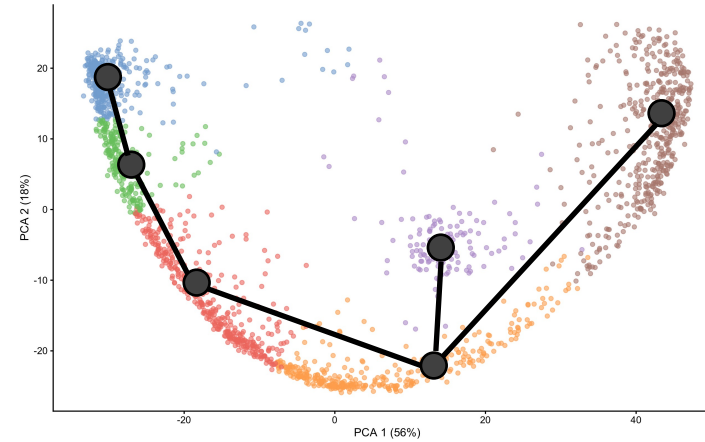
CAREFUL:

Once again, don't get tricked by the 2D visualization... Here, the principal curve is computed from 50 PCs, and subsequently embedded in only 2 PCs.



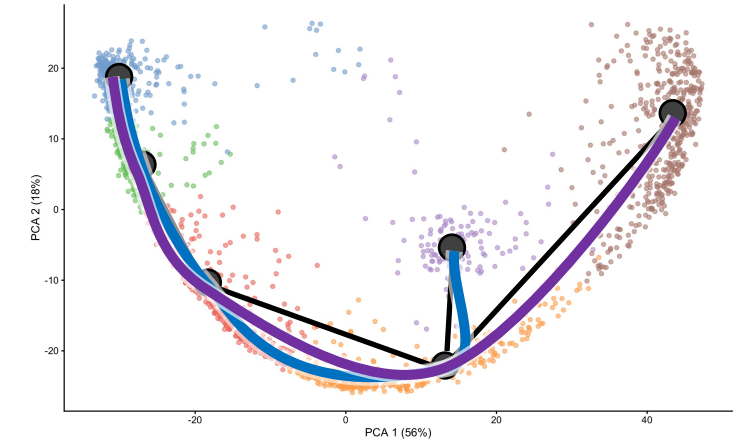
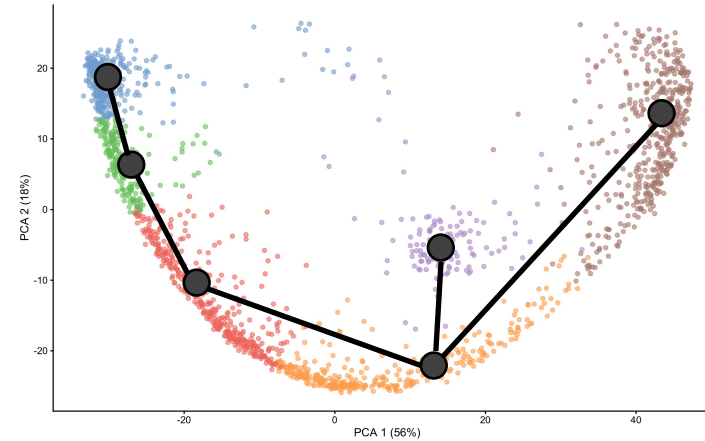
Why clustering quality matters for trajectory inference

The MST is built from cluster centroids. If clustering changes, the MST will change and the fitted principal curves too.

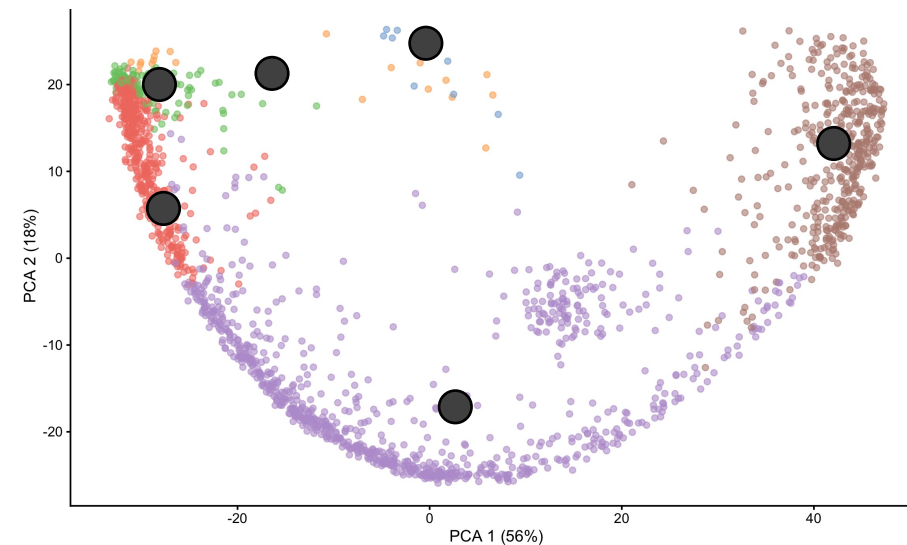


Why clustering quality matters for trajectory inference

The MST is built from cluster centroids. If clustering changes, the MST will change and the fitted principal curves too.



Only perform TI once you are confident your clusters represent a biologically relevant cell population.



Dynverse project provides a handy exploratory/summarizing app to pick the best-suited TI algo.

▼ Topology DEFAULT

Do you expect multiple disconnected trajectories in the data?

Yes
I don't know
No

▼ Scalability COMPUTED

Number of cells

1000

Number of features (genes)

1000

Time limit

10s

1h

∞

Memory limit

100MB

30GB

∞

▼ Prior information DEFAULT

Are you able to provide the following prior information?

Start cell(s), End cell(s), # end states, # start states, # leaves, # states, Marker genes, A dii ▼

► Method selection DEFAULT

► Benchmarking metrics DEFAULT

► Availability COMPUTED

► Benchmarking datasets DEFAULT

Show code `</>`

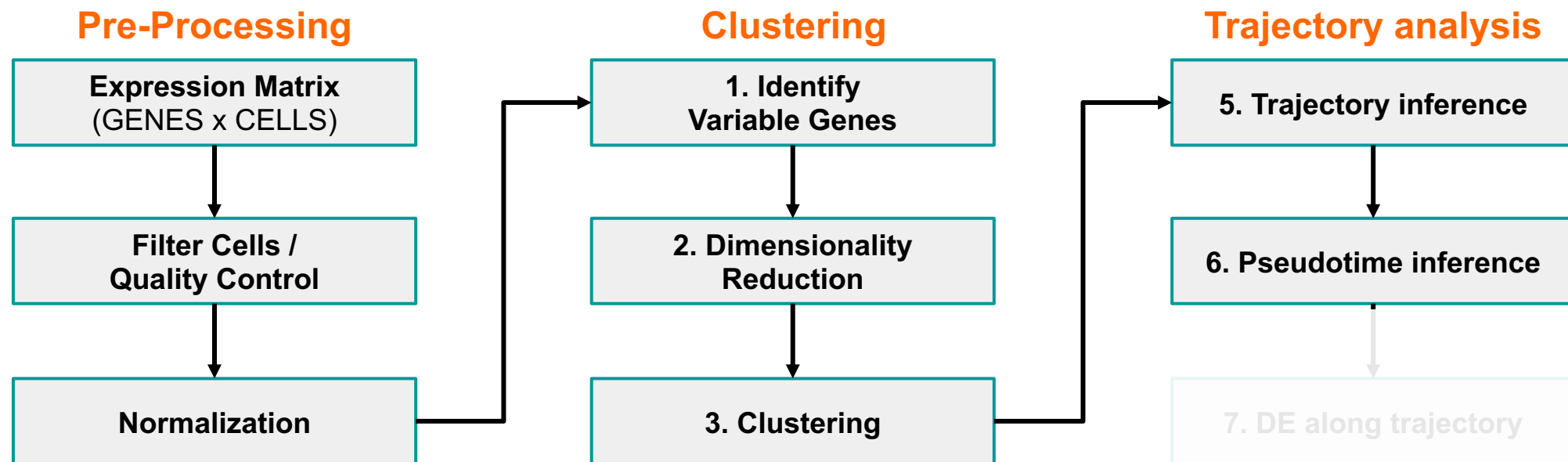
Show/hide columns `⌵`

Options `⚙`

Infer trajectories with *dynamo* `↻`

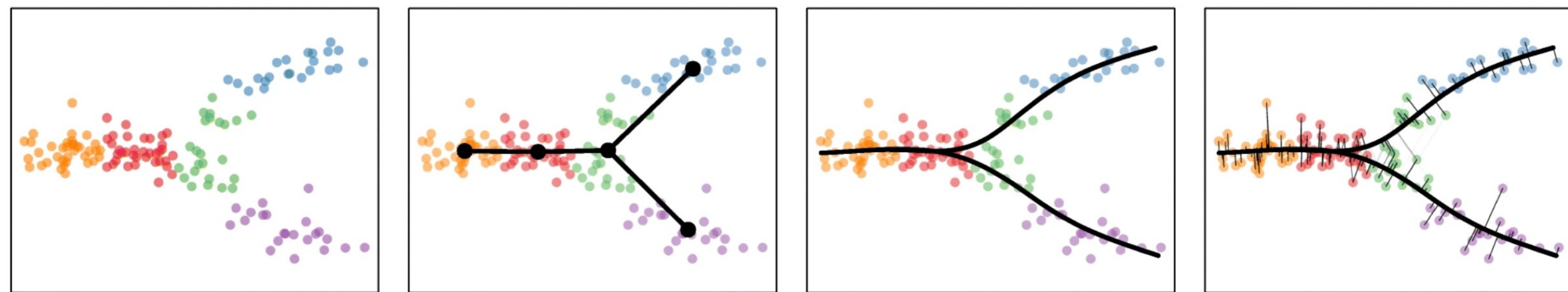
Lenses	Default	Summary (Fig. 2)	Method	Scalability	Stability	Usability	Accuracy	Overall	Everything	
Method								Accuracy	Scalability	Stability
✓ <code></></code>	🚀 Name		Priors		Errors		Overall IF	🕒	📀	Stability
✓	🚀 Slingshot						100	8s	942MB	
✓	🚀 PAGA Tree	×					99	19s	625MB	⚠️ Unstable
✓	🚀 SCORPIUS						96	3s	507MB	
✓	Angle						92	1s	308MB	
	🚀 PAGA	×					89	15s	559MB	⚠️ Unstable
	🚀 Embeddr						89	5s	591MB	
	MST						89	4s	572MB	⚠️ Unstable
	🚀 Waterfall						89	5s	369MB	
	🚀 TSCAN						88	5s	476MB	⚠️ Unstable
	Component 1						87	1s	516MB	
	🚀 SLICE						83	16s	713MB	
	🚀 EIPiGraph linear						81	1m	573MB	
	🚀 PhenoPath						79	5m	837MB	
	🚀 pCrestode						78	2m	444MB	⚠️ Unstable
	🚀 Monocle ICA	×					78	1m	692MB	⚠️ Unstable
	🚀 Wanderlust	×					78	51s	413MB	
	🚀 MATCHER						77	43s	385MB	
	🚀 Wishbone	×					76	1m	370MB	
	🚀 EIPiGraph cycle						76	1m	532MB	⚠️ Unstable
	🚀 cellTree maptpx						74	4m	692MB	⚠️ Unstable
	🚀 MFA	×					72	7m	669MB	
	🚀 SCUBA						70	5m	418MB	⚠️ Unstable

Analysis workflow



Pseudotime inference

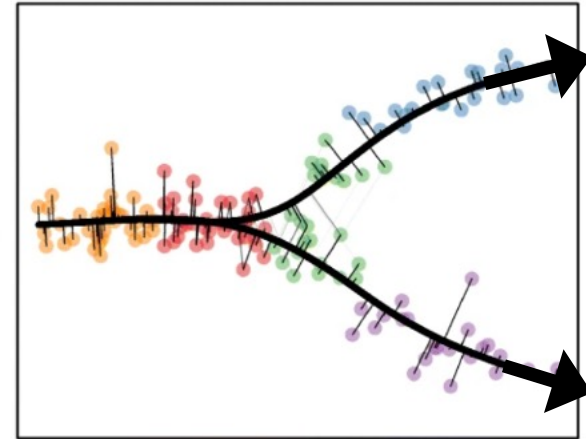
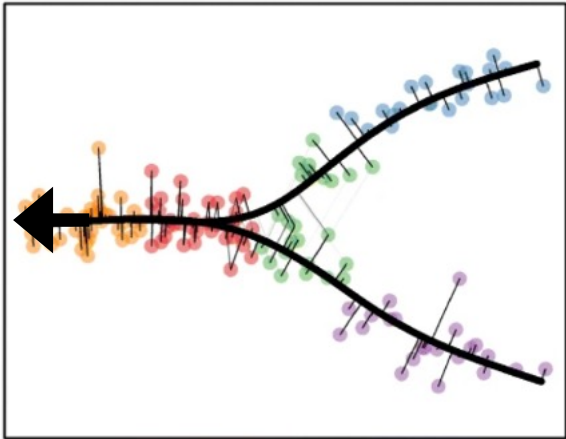
In Slingshot, the pseudotimes values are estimated as the euclidean distance from each point to the closest principal curve (i.e. by **orthogonal projection of each point onto the curve**).



But how are trajectories oriented???

A trajectory is not a vector: a trajectory is not oriented!

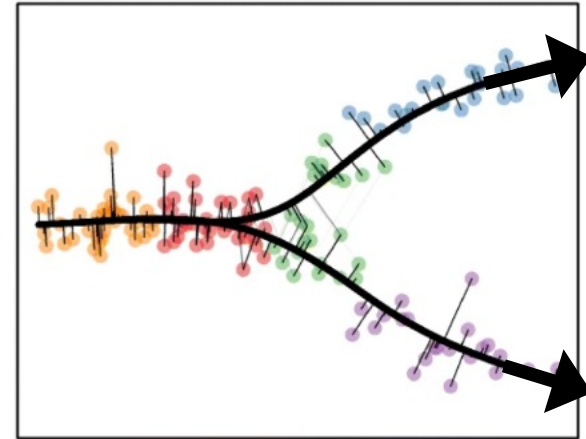
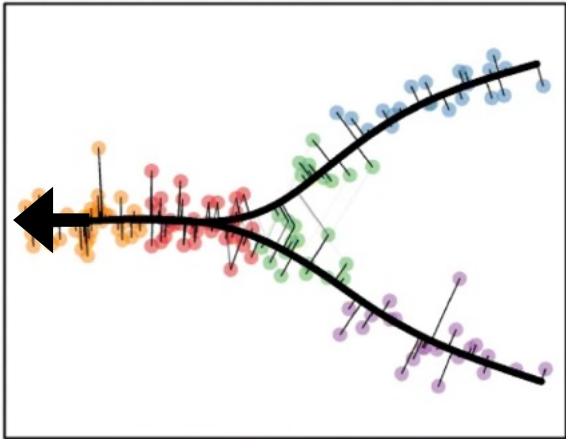
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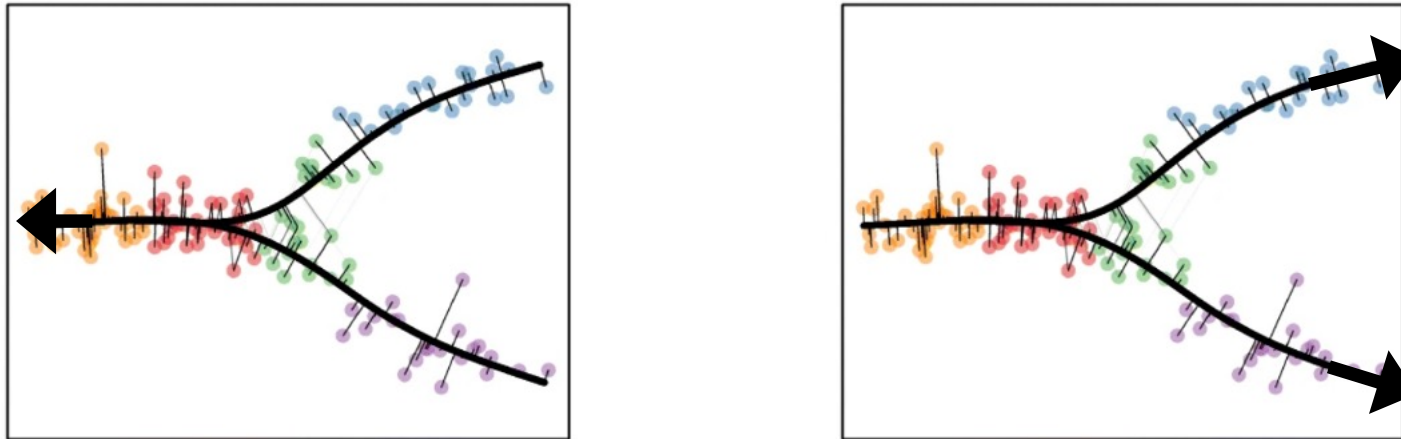


That is when one should use prior knowledge, ground-truth, or simply make a call.

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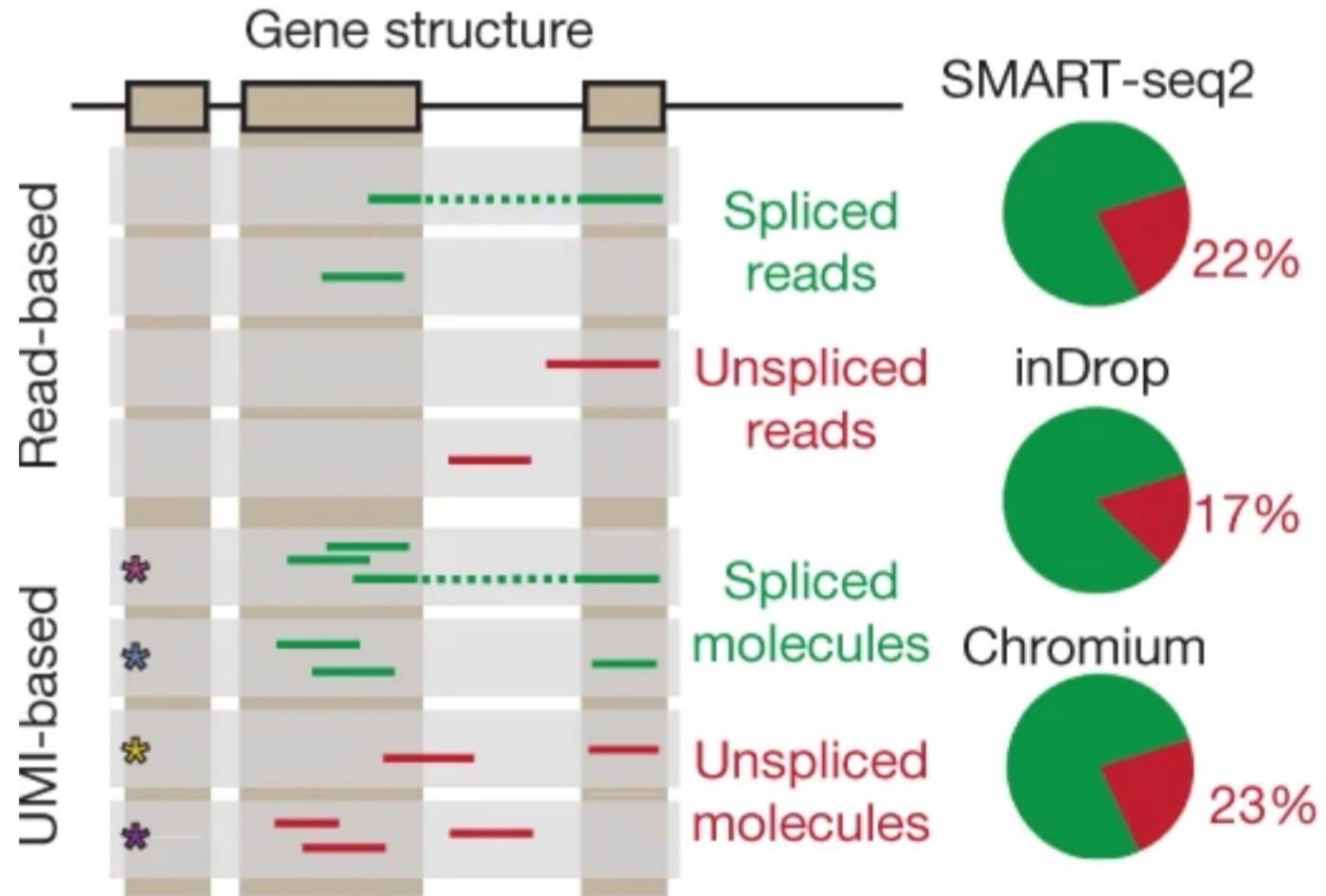
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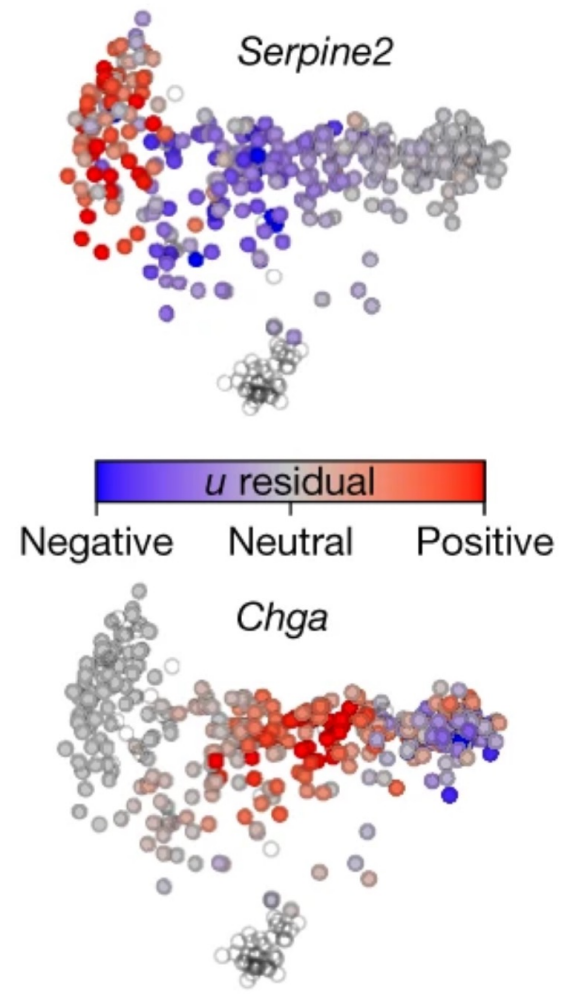
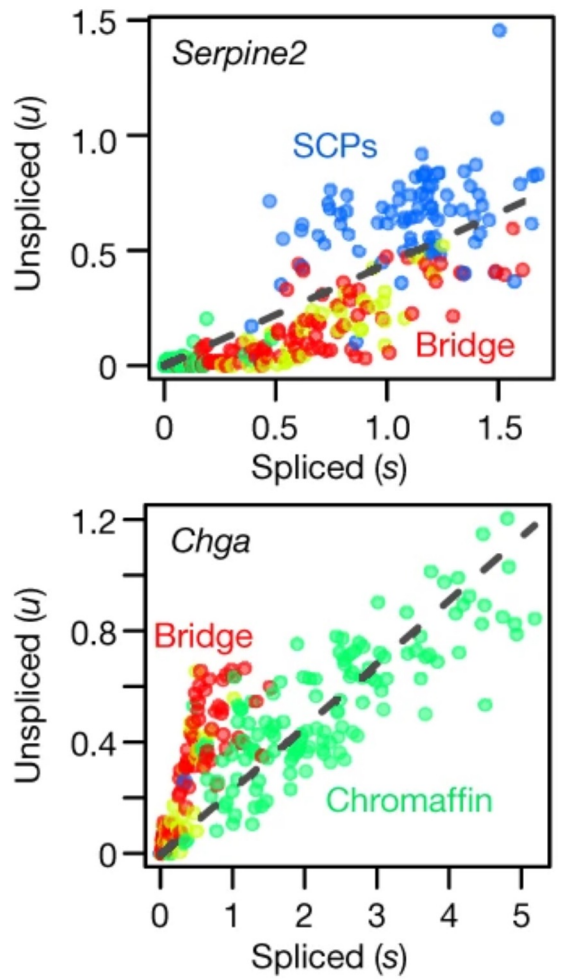
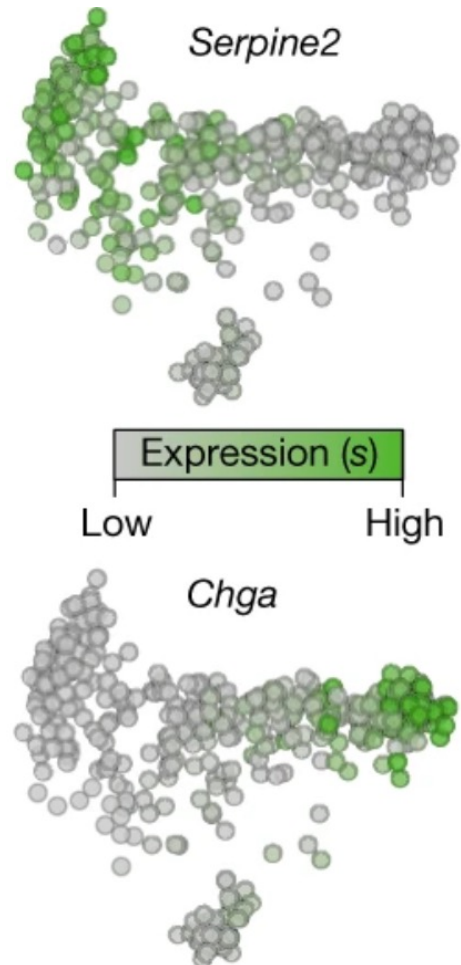
- Hematopoietic stem cell
- Lymphoid stem cell
- Committed lymphoid stem cell
- B cell
- T cell

RNA velocity

- Single-cell RNA-seq reads can be mapped onto exons or introns.

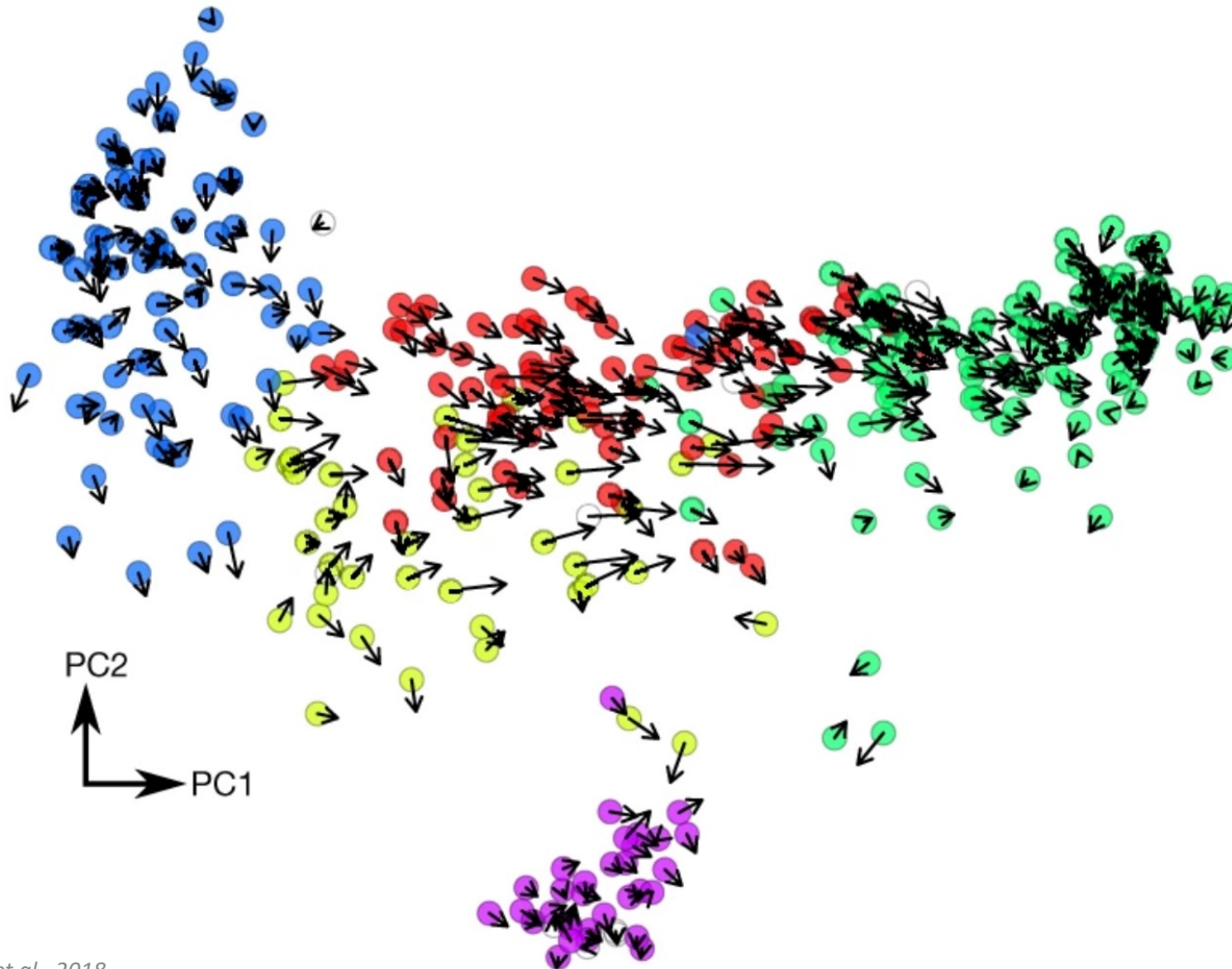


RNA velocity

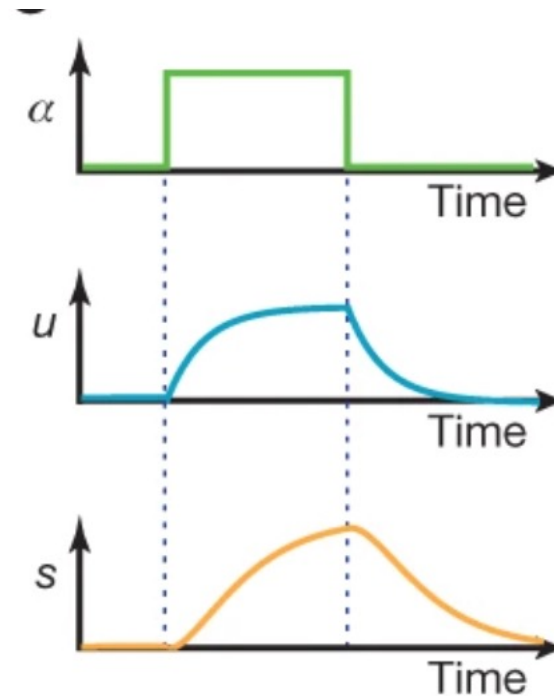
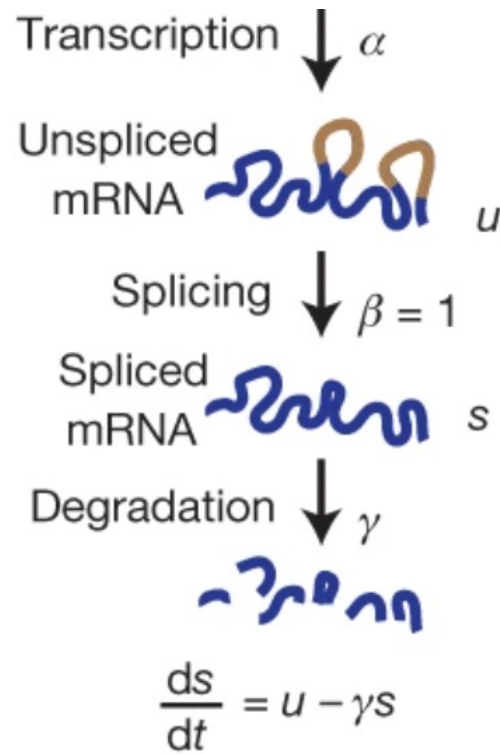


u residual represents whether the cell is far from an equilibrium state of expression

Generalizing this to all the detected (variable) genes, one can infer the future "position" of each cell.



Generalizing this to all the detected (variable) genes, one can infer the future "position" of each cell.



CAREFUL!!!

This model relies on an important assumption:

that transcription is steady-state.

This is rarely true, even less true for differentiating cells!!

Article | Published: 03 August 2020

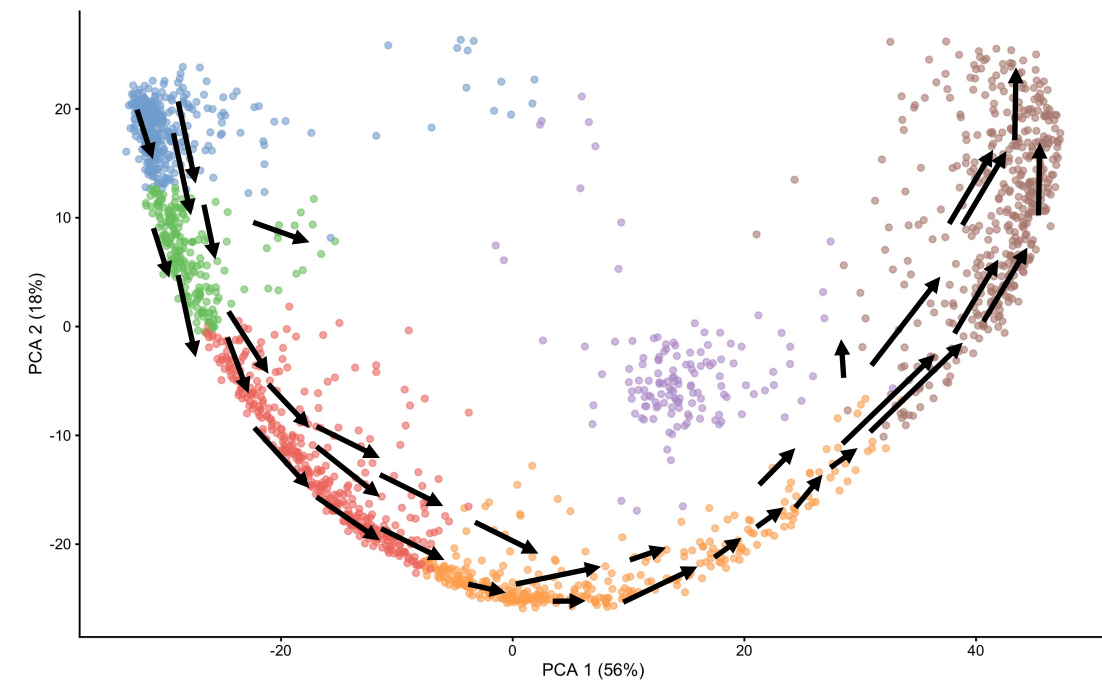
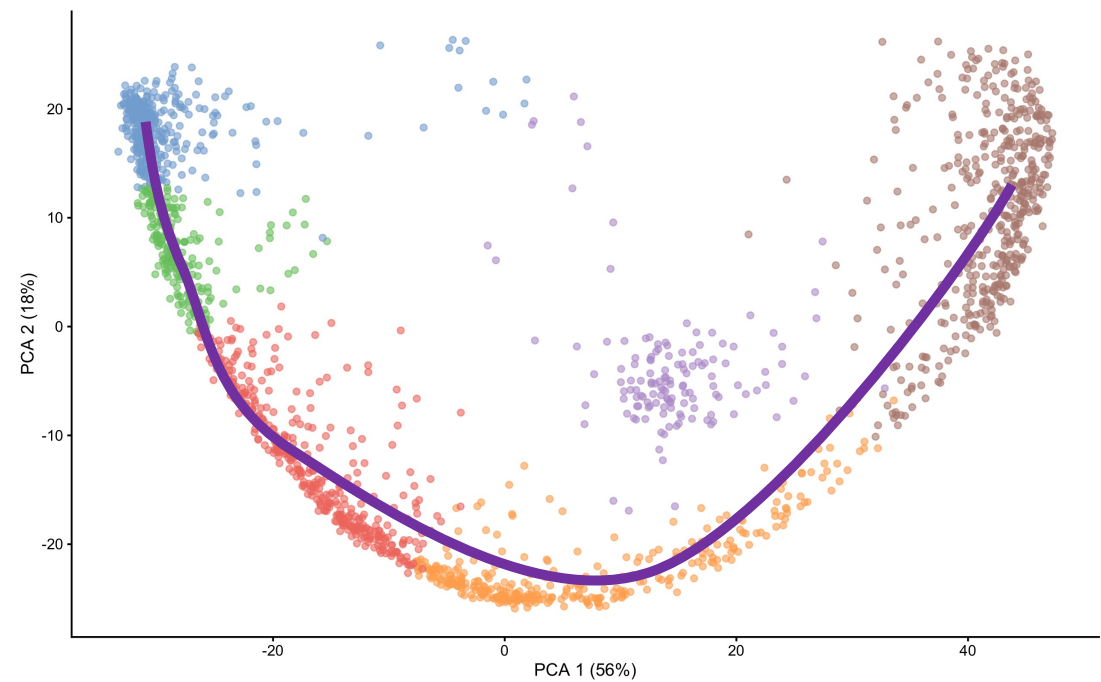
Generalizing RNA velocity to transient cell states through dynamical modeling

Volker Bergen, Marius Lange, Stefan Peidli, F. Alexander Wolf  & Fabian J. Theis 

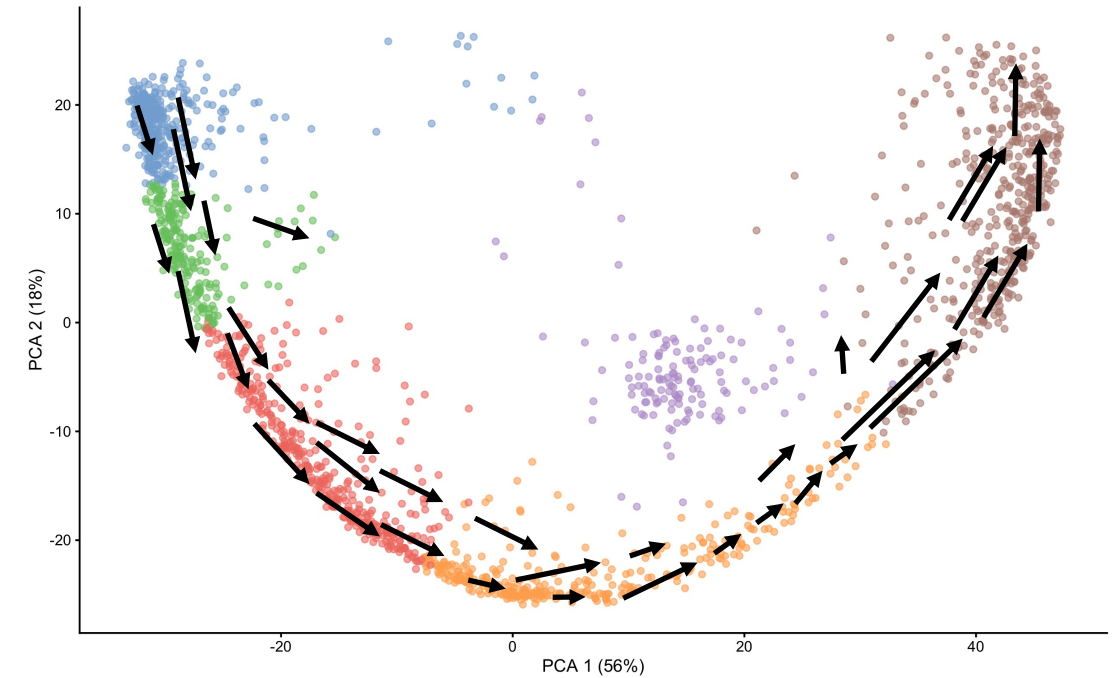
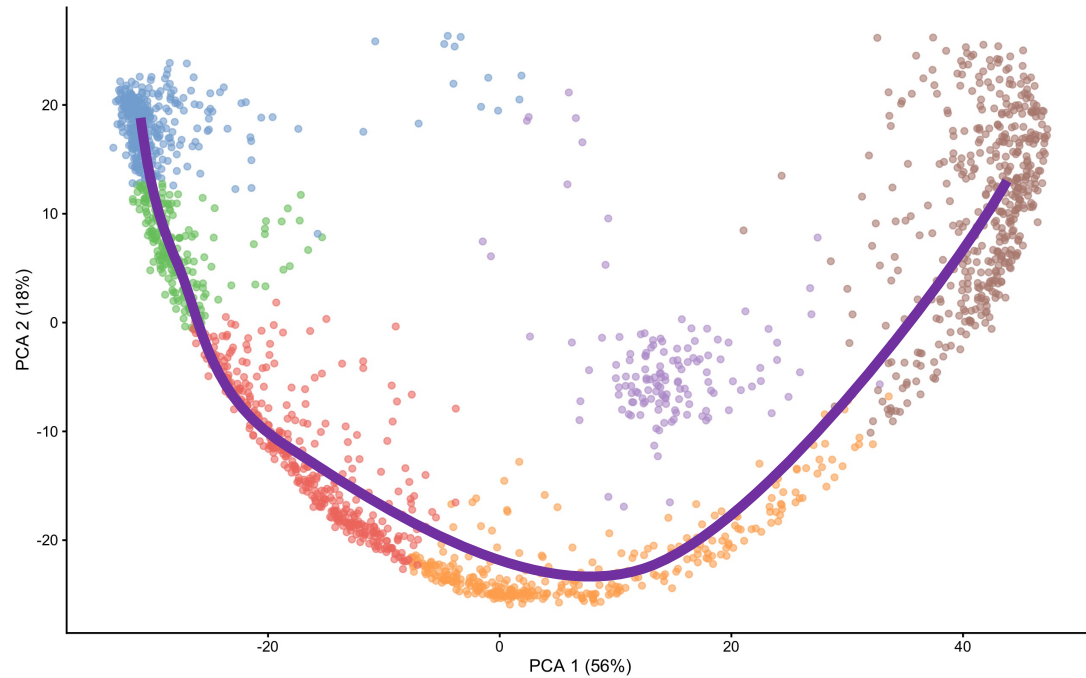
Nature Biotechnology **38**, 1408–1414 (2020) | [Cite this article](#)

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Using RNA velocity to infer directionality of the trajectory

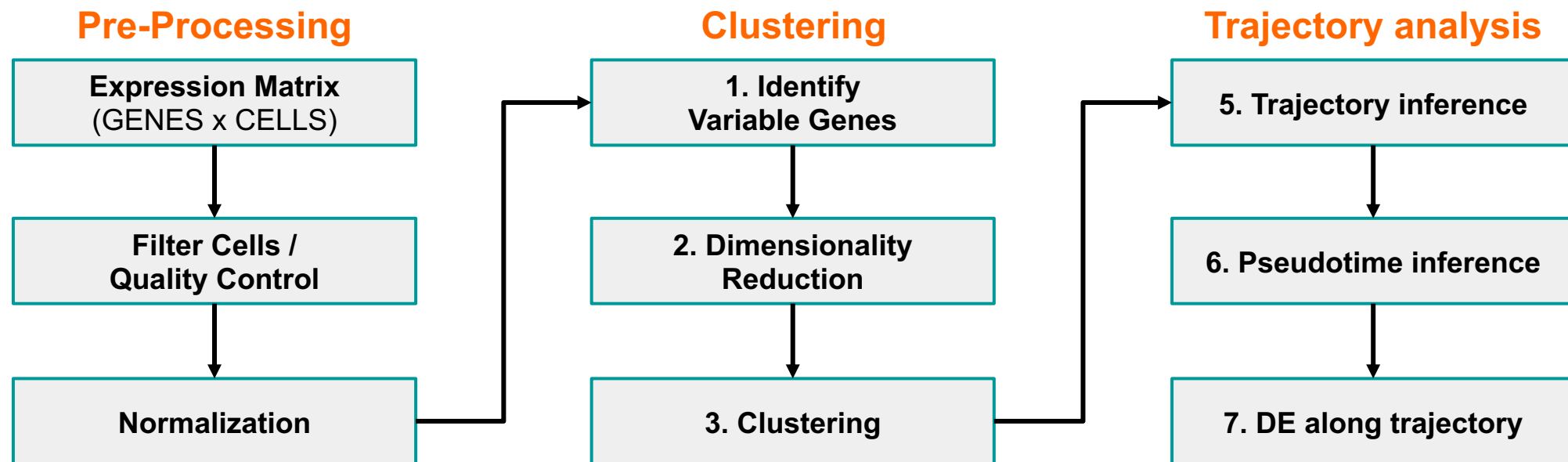


Using RNA velocity to infer directionality of the trajectory



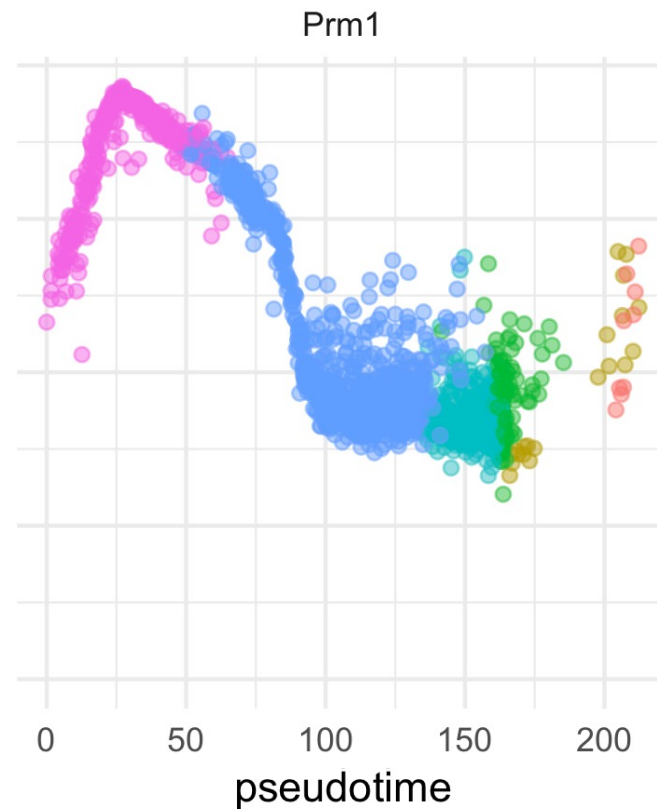
RNA velocity implementations generally output a pseudotime value. However, they do not infer trajectories!! So they cannot capture a branching event, or cycles, etc....

Analysis workflow



Modeling gene expression along pseudotime

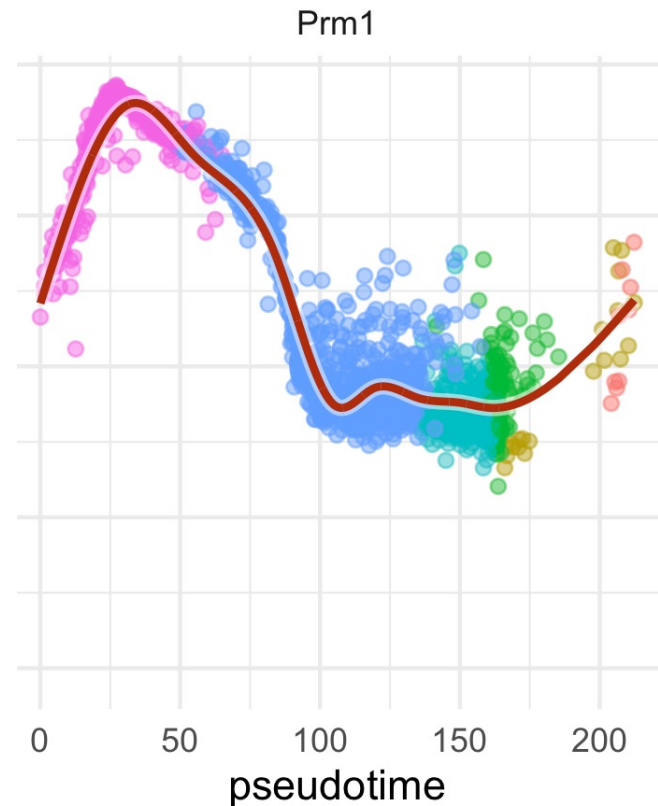
With a pseudotime value inferred to each cell, one can place all cells along an x axis, and plot gene expression on an y axis.



Modeling gene expression along pseudotime

With a pseudotime value inferred to each cell, one can place all cells along an x axis, and plot gene expression on an y axis.

Then time-dependent expression can be modelled, typically by fitting a GAM to the gene expression ~ pseudotime.



tradeSeq is an R package which facilitates GAM-fitting and gene DE analysis along and between trajectories.

It implements plug-and-play methods to use outputs from Slingshot, but most functions can be used directly with pseudotime values, without having to rely on trajectories specifically inferred with Slingshot.

Article | [Open Access](#) | Published: 05 March 2020

Trajectory-based differential expression analysis for single-cell sequencing data

Koen Van den Berge, Hector Roux de Bézieux, Kelly Street, Wouter Saelens, Robrecht Cannoodt, Yvan Saeys, Sandrine Dudoit & Lieven Clement

Nature Communications 11, Article number: 1201 (2020) | [Cite this article](#)

32k Accesses | 24 Citations | 65 Altmetric | [Metrics](#)

