

Lecture 6

Inferring cell pseudotime from scRNAseq data

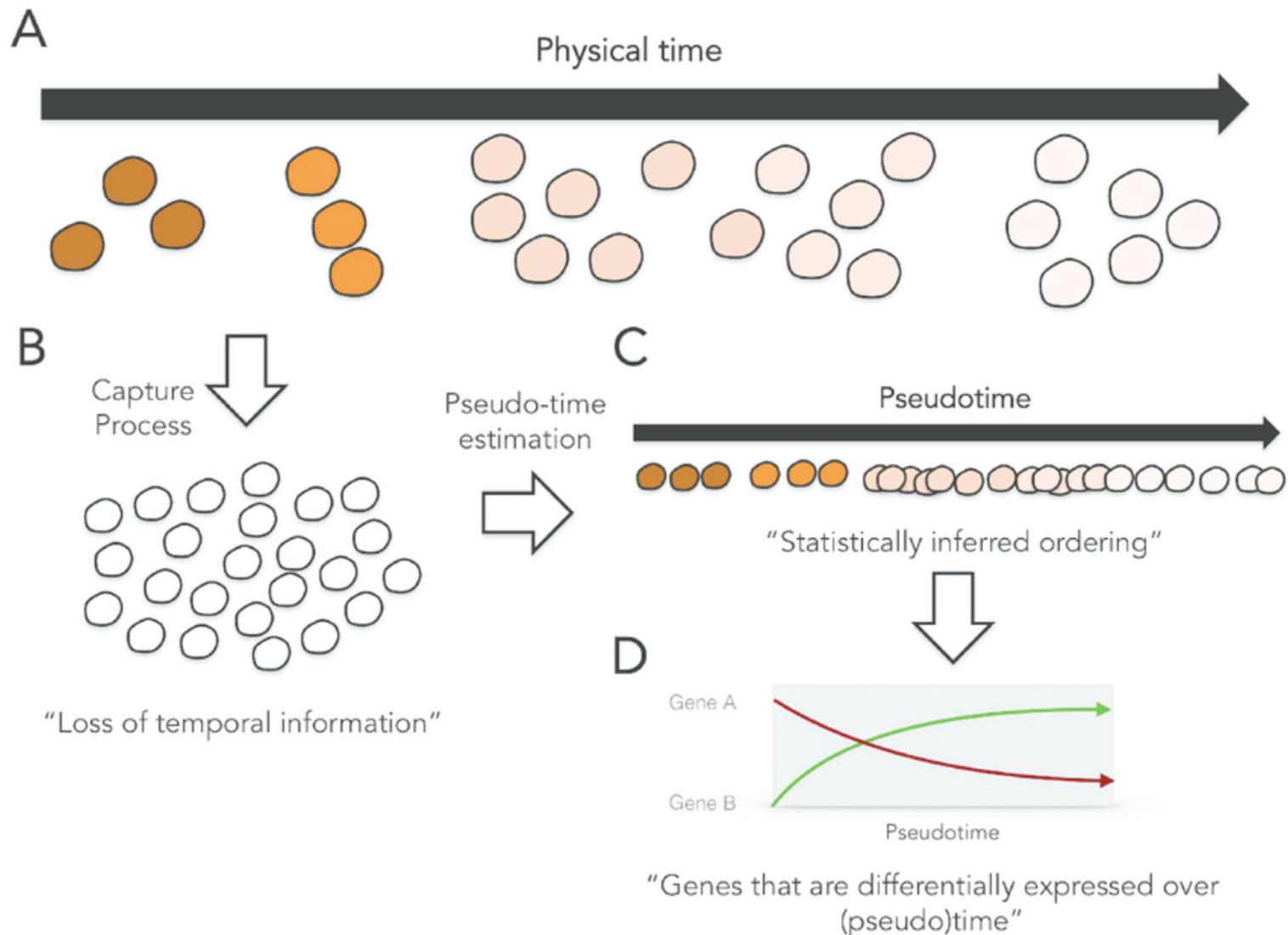
Physalia course 2023

—

Single-cell RNA-seq with R/Bioconductor

Instructors: Orr Ashenberg & Jacques Serizay

What is pseudotime?



What is pseudotime?



- Pseudotime is an abstract unit of progress: it's simply the distance between a cell and the start of the trajectory, measured along the shortest path
- The trajectory's total length is defined in terms of the total amount of transcriptional change that a cell undergoes as it moves from the starting state to the end state.
- **Pseudotime is a measure of how much progress an individual cell has made through a process (such as cell differentiation).**

What is pseudotime in single-cell RNA-seq?

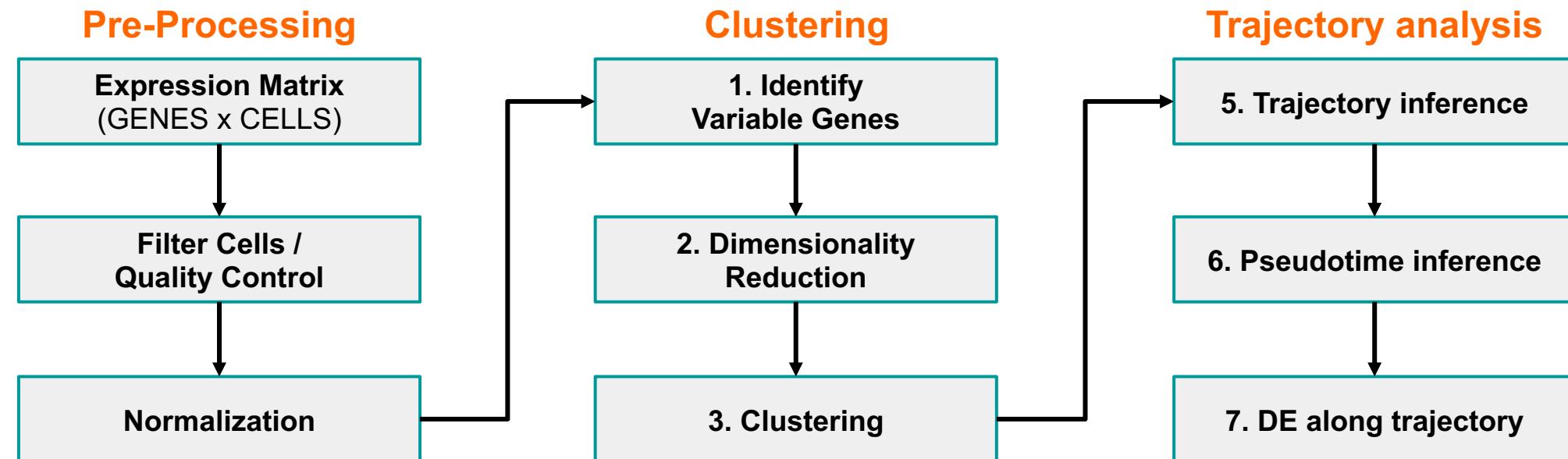
- In single-cell expression studies of processes such as cell differentiation, captured cells might be widely distributed in terms of progress.
- That is, in a population of cells captured at exactly the same time, some cells might be far along, while others might not yet even have begun the process.
- **By ordering each cell according to its progress along a learned trajectory, pseudotime inference alleviates the problems that arise due to asynchrony.**

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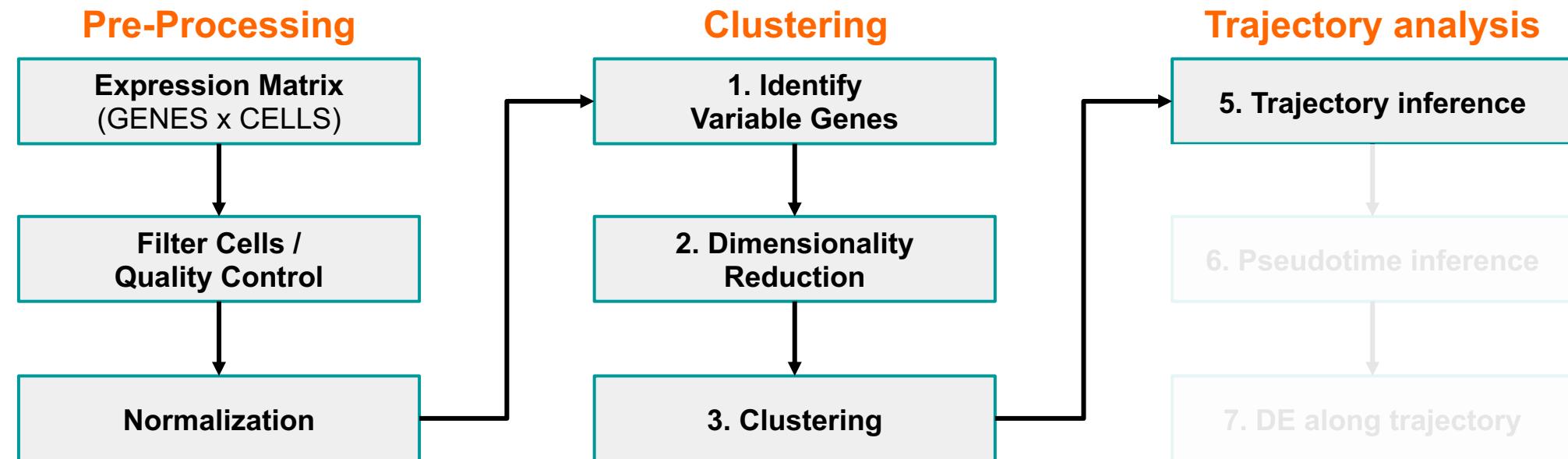
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This is why pseudotime and trajectory inference are largely overlapping.

Analysis workflow

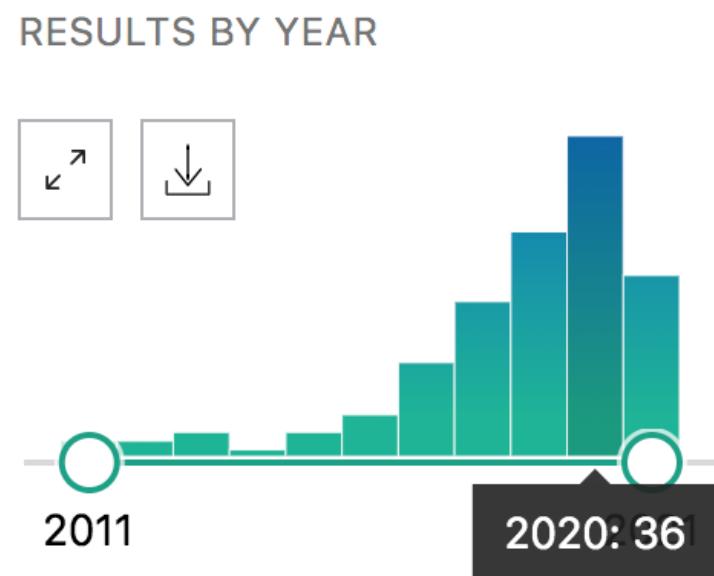


Analysis workflow



A new algorithm comes out ~ every other week...

Pubmed results for “**trajectory inference method single-cell**”:



A new algorithm comes out ~ every other week...

Pseudocell Tracer-A **method for inferring dynamic trajectories** using scRNASeq and its application to B cells undergoing immunoglobulin class switch recombination.

Reiman D, Manakkat Vijay GK, Xu H, Sonin A, Chen D, Salomonis N, Singh H, Khan AA.

PLoS Comput Biol. 2021 May 3;17(5):e1008094. doi: 10.1371/journal.pcbi.1008094. eCollection 2021

May.

PMID: 33939691 **Free PMC article.**

Single cell RNA sequencing (scRNASeq) can be used to **infer** a temporal ordering of cellular states.

Current **methods for the inference of cellular trajectories** rely on unbiased dimensionality reduction techniques. ...

Slingshot: cell lineage and pseudotime inference for single-cell transcriptomics.

Street K, Risso D, Fletcher RB, Das D, Ngai J, Yosef N, Purdom E, Dudoit S.

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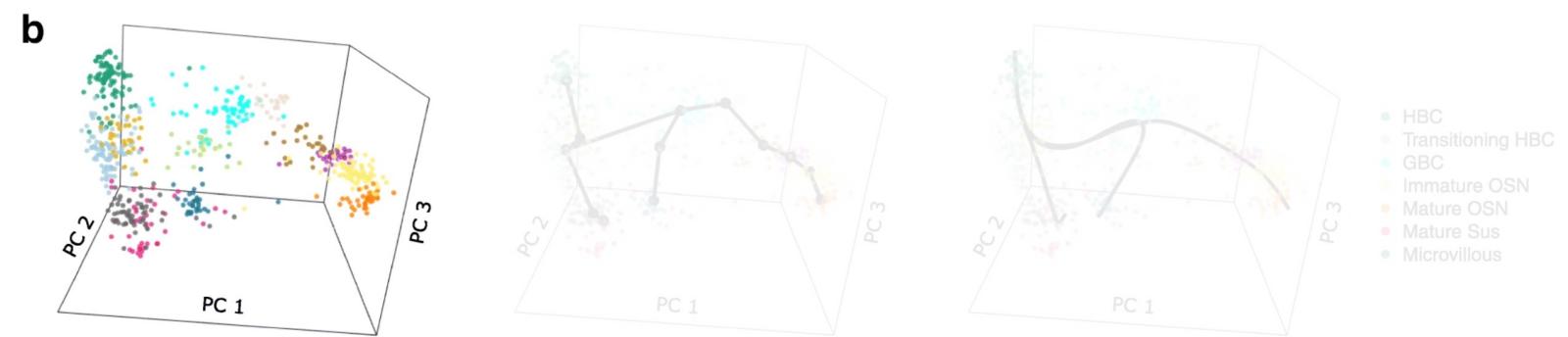
Stick to **standards!!**

QCed, troubleshooted, optimized and **generic**

Slingshot is one of the most widely used and robust approaches to infer trajectory in relatively simple datasets.

It works by:

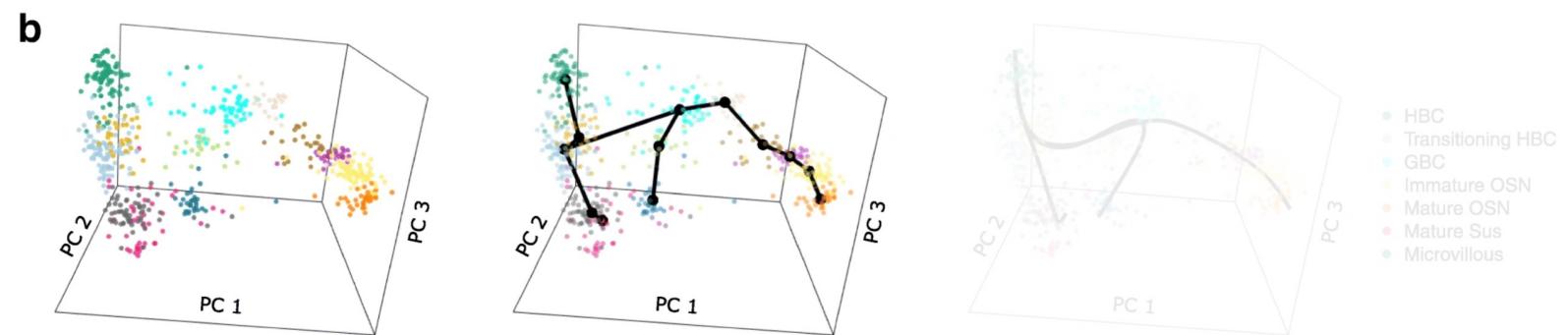
0. Get clustered data in a low-dimensional space



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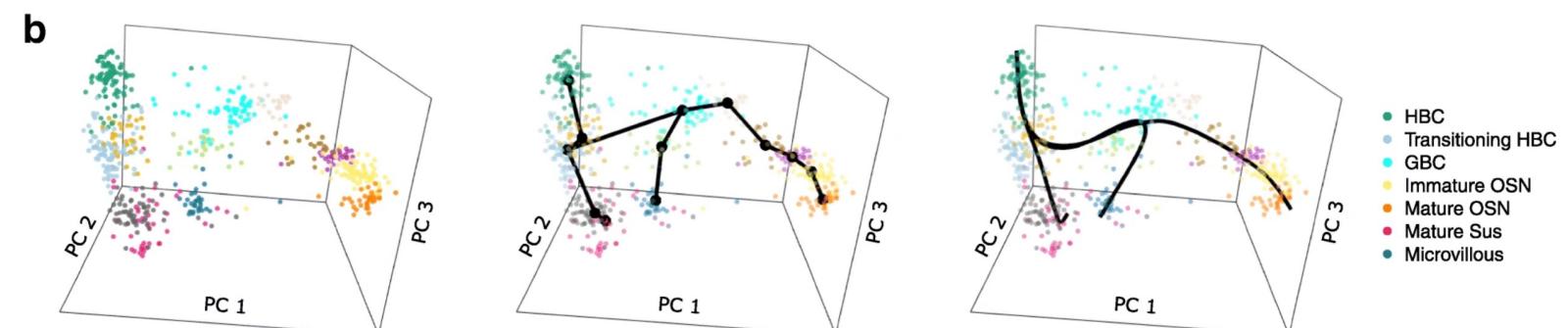
0. Get clustered data in a low-dimensional space
1. Building a minimum spanning tree on the clusters



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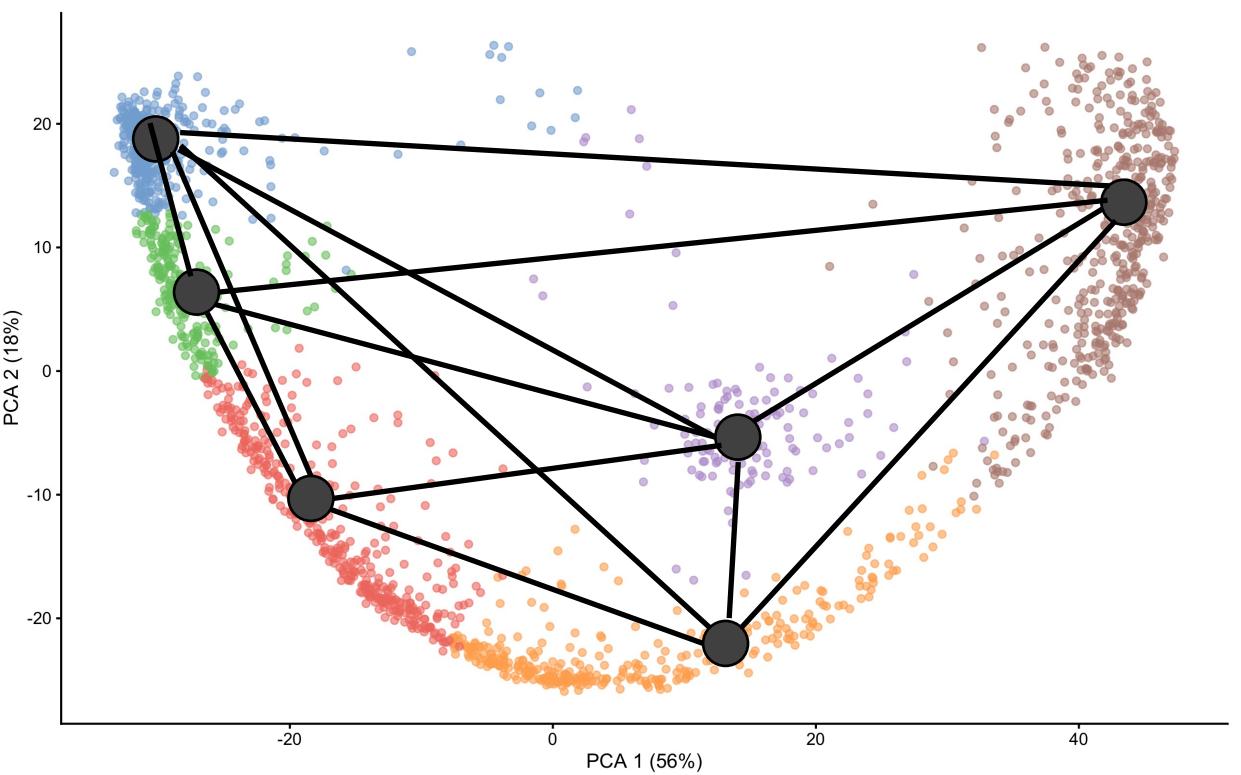
0. Get clustered data in a low-dimensional space
1. Building a minimum spanning tree on the clusters
2. Fit principal curves through the MST



Minimum spanning tree?!

Or minimum weight spanning tree

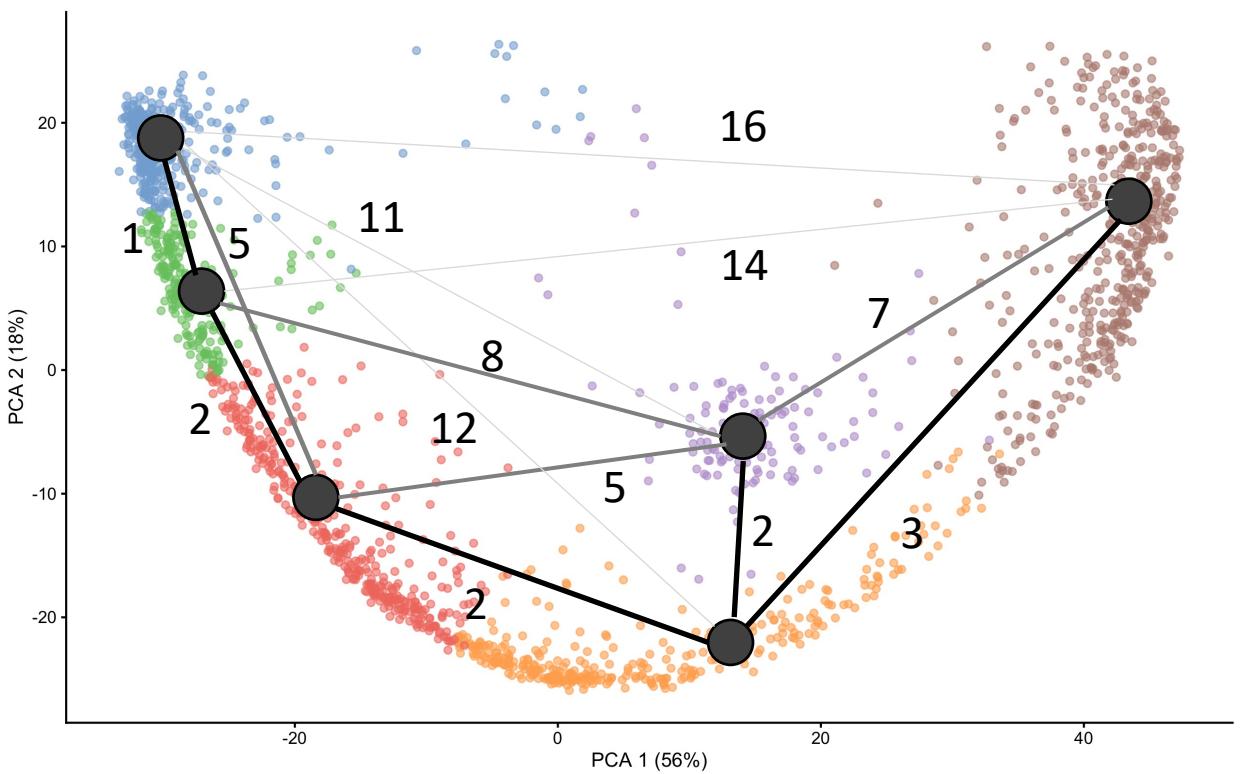
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Minimum spanning tree?!

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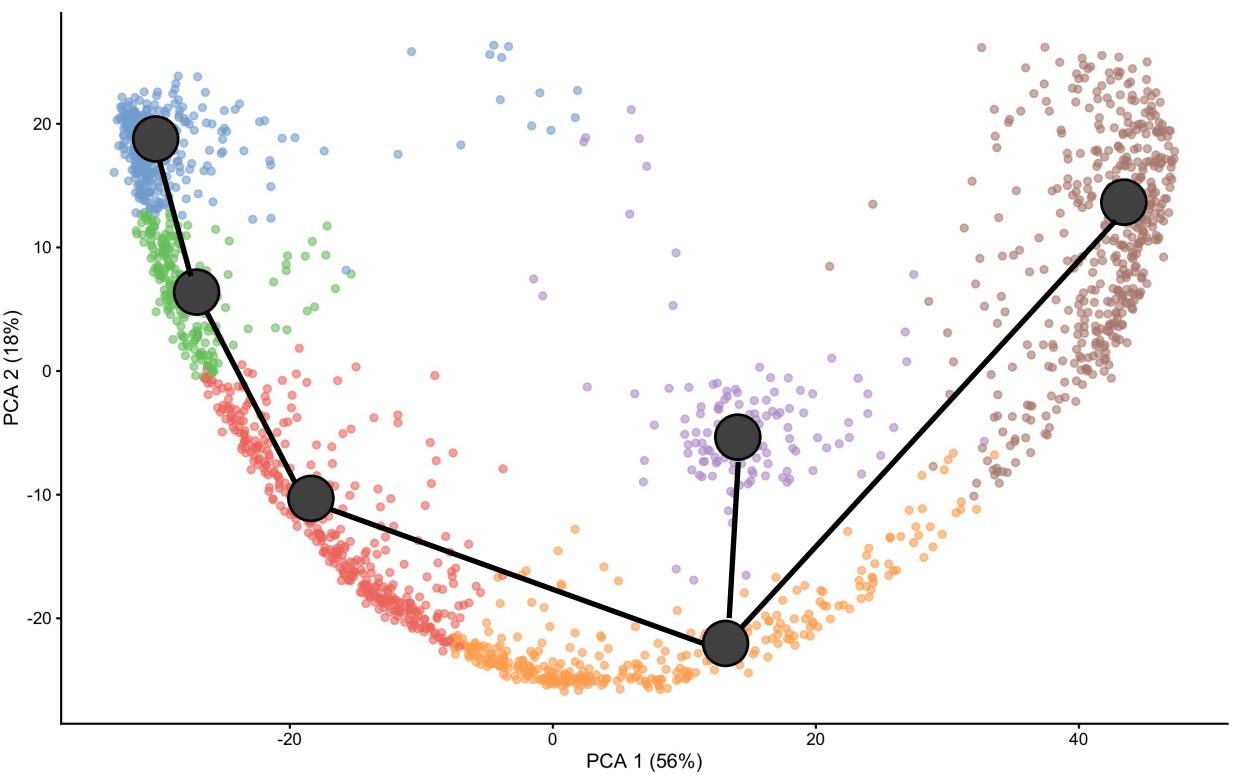
1. Build a graph with edges between each pair of clusters
2. Add weights to each edge according to proximity of the two clusters



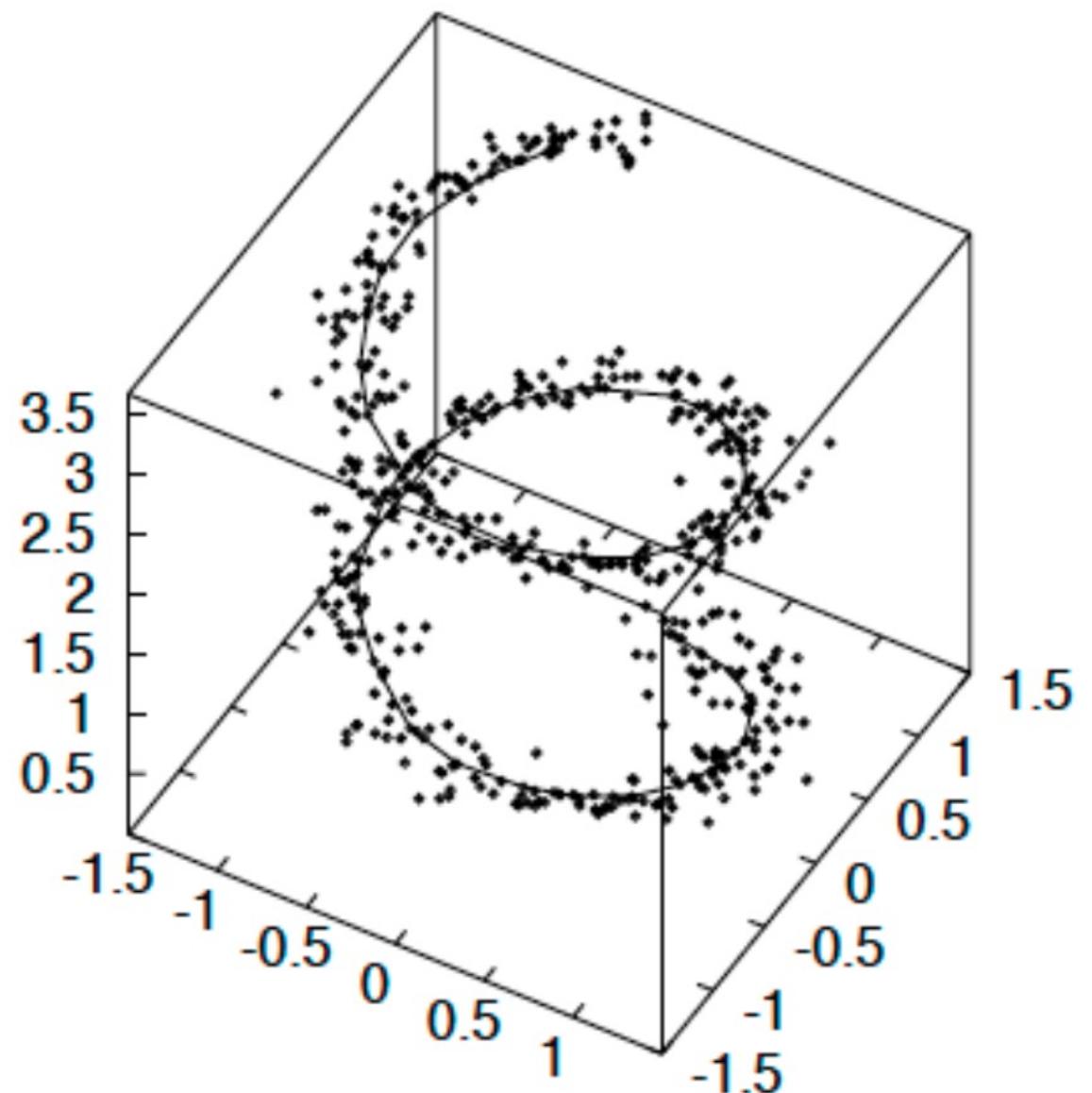
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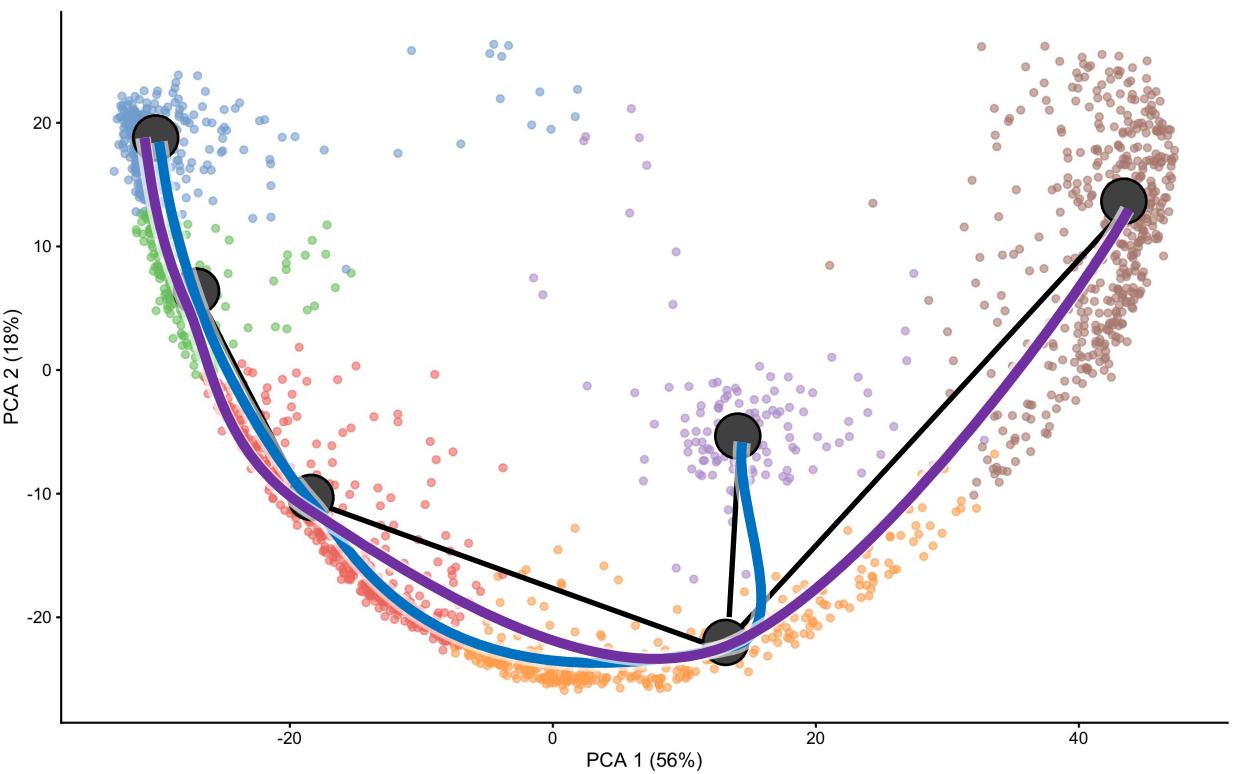
1. Build a graph with edges between each pair of clusters
2. Add weights to each edge according to proximity of the two clusters
3. Find the shortest path between clusters



A principal curve is a smooth, one-dimensional, curve that passes through the middle of a high-dimensional data set, providing a nonlinear summary of the data.



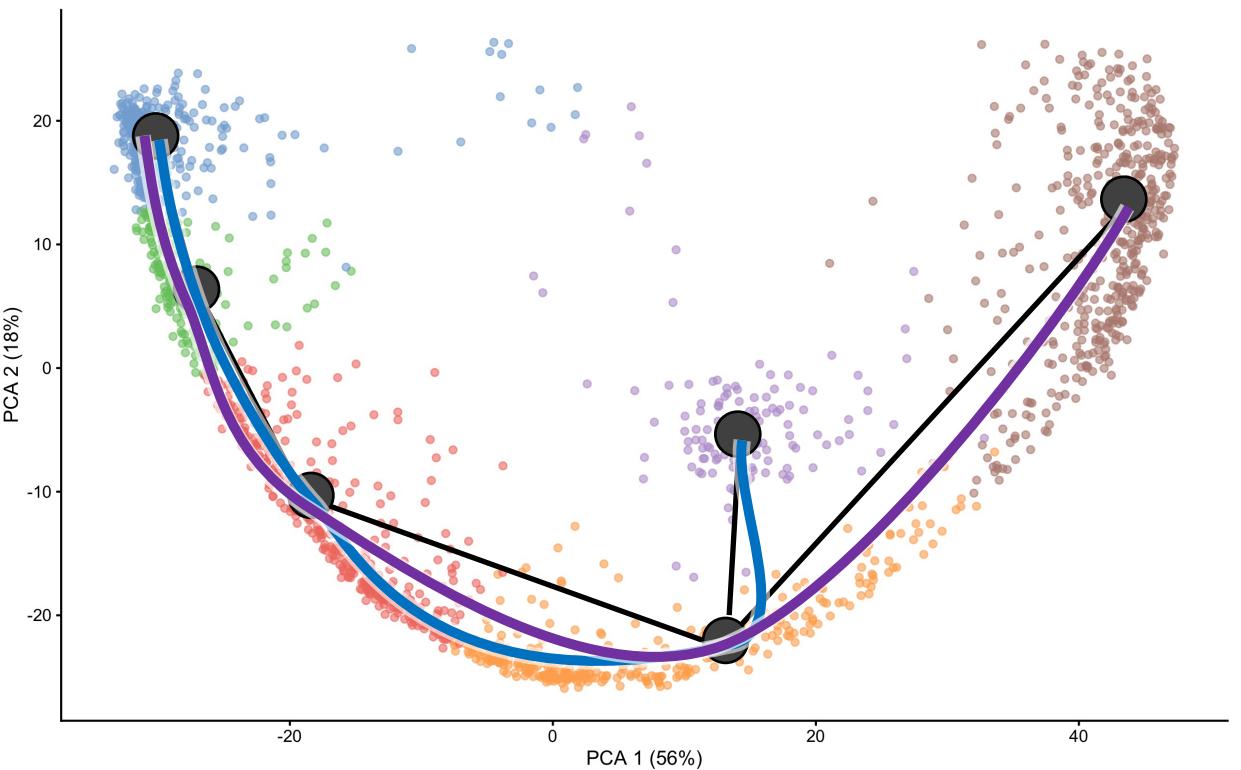
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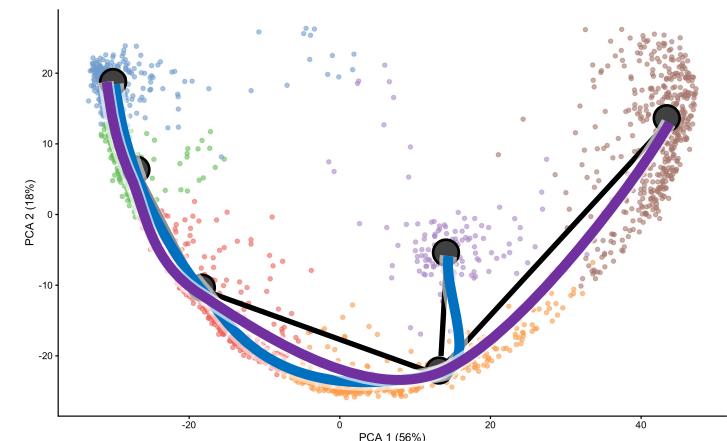
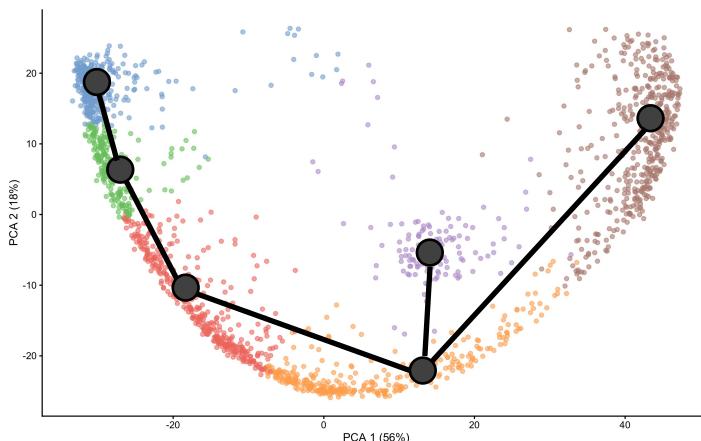
CAREFUL:

Once again, don't get tricked by the 2D visualization... Here, the principal curve is computed from 50 PCs, and subsequently embedded in only 2 PCs.



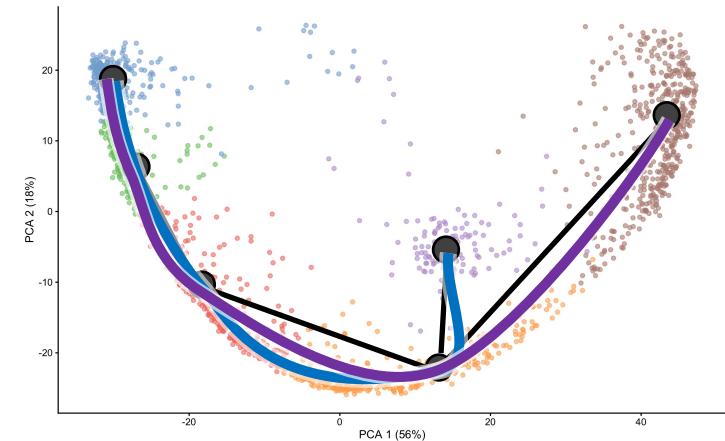
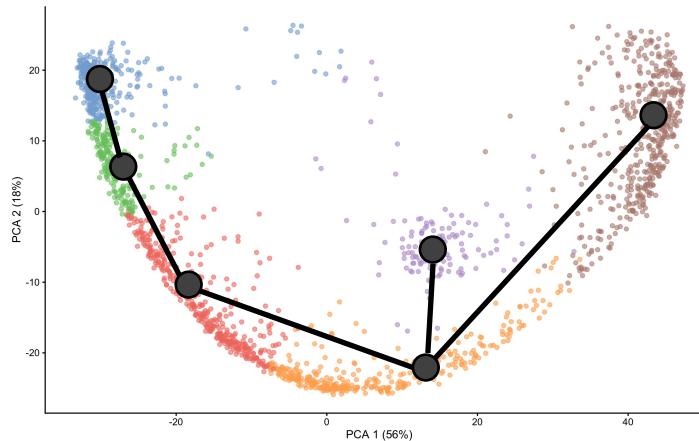
Why clustering quality matters for trajectory inference

The MST is built from cluster centroids. If clustering changes, the MST will change and the fitted principal curves too.

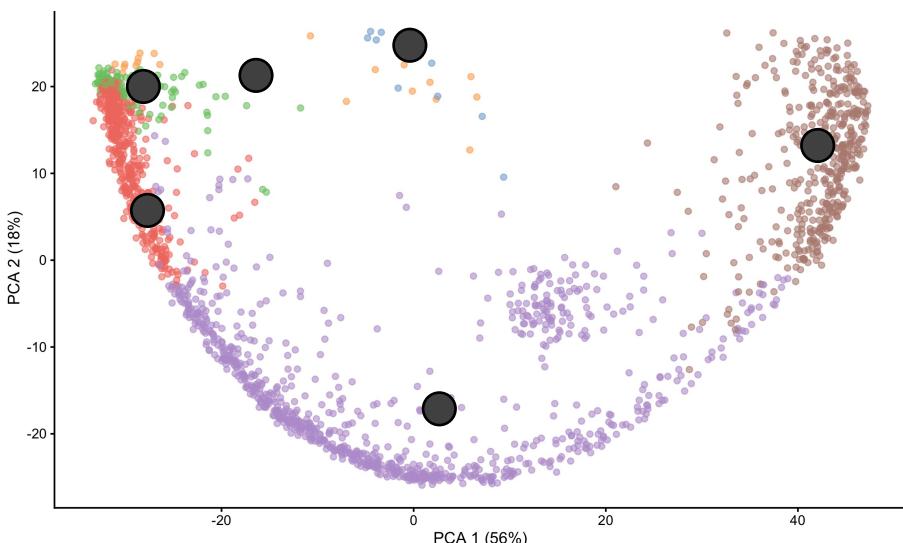


Why clustering quality matters for trajectory inference

The MST is built from cluster centroids. If clustering changes, the MST will change and the fitted principal curves too.



Only perform TI once you are confident your clusters represent a biologically relevant cell population.



Making sense of all these different methods



Dynverse project provides a handy exploratory/summarizing app to pick the best-suited TI algo.

Topology

Scalability

Number of cells: 1000

Number of features (genes): 1000

Time limit: 10s to 1h

Memory limit: 100MB to 30GB

Prior information

Are you able to provide the following prior information?

Start cell(s), End cell(s), # end states, # start states, # leaves, # states, Marker genes, A di

Method selection

Benchmarking metrics

Availability

Benchmarking datasets

Show code ↕

Show/hide columns ⓘ

Options ⚙

Infer trajectories with dyno ↗

Lenses: Default, Summary (Fig. 2)

Method

Scalability

Stability

Usability

Accuracy

Overall

Everything

Method

Priors

Errors

Overall

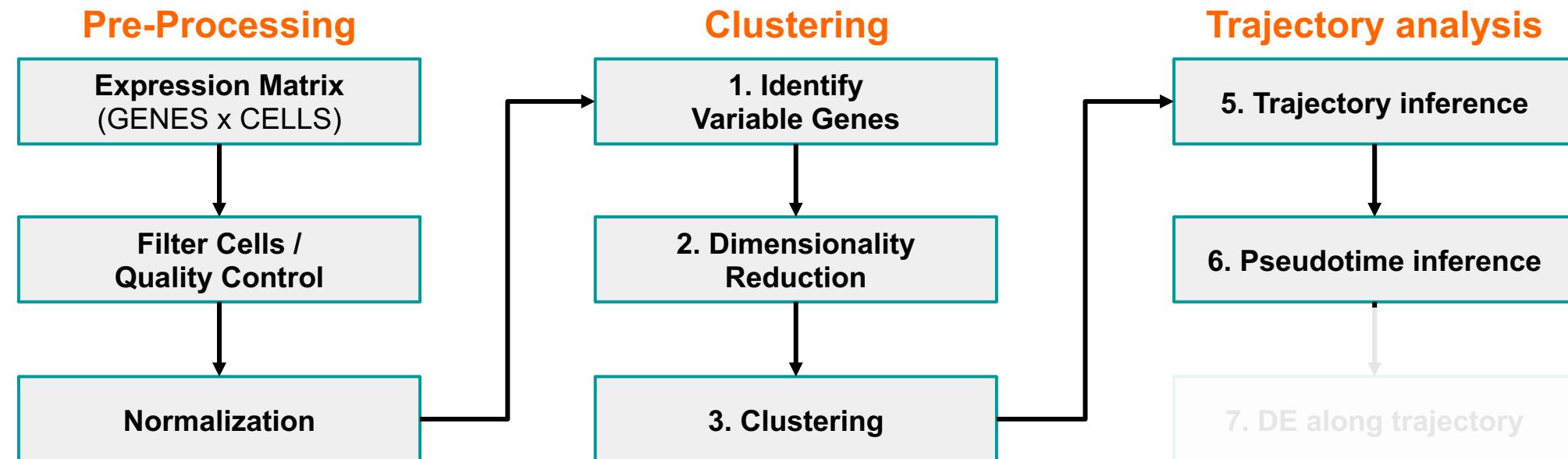
Accuracy

Scalability

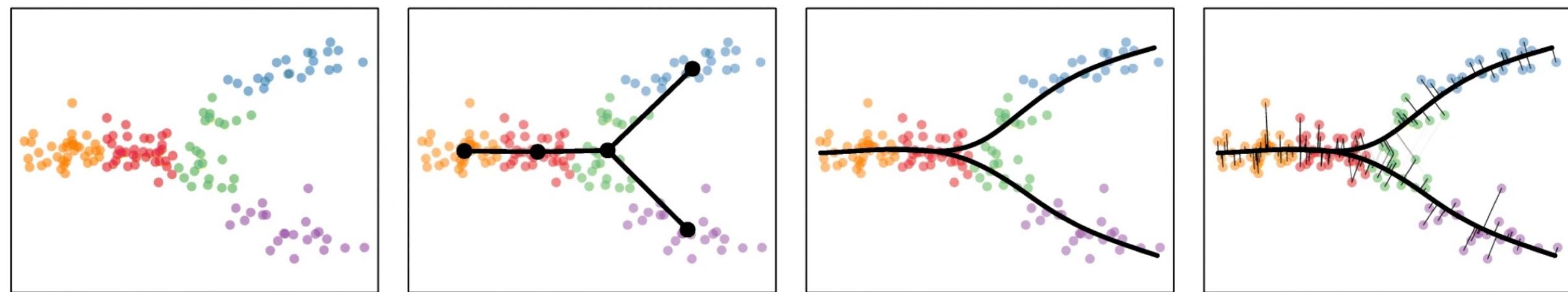
Stability

Method	Priors	Errors	Overall	Accuracy	Scalability	Stability
Slingshot	✗	100	8s	942MB		
PAGA Tree	✗	99	19s	625MB	⚠️ Unstable	
SCORPIUS	✗	96	3s	507MB		
Angle	✗	92	1s	308MB		
PAGA	✗	89	15s	559MB	⚠️ Unstable	
Embeddr	✗	89	5s	591MB		
MST	✗	89	4s	572MB	⚠️ Unstable	
Waterfall	✗	89	5s	369MB		
TSCAN	✗	88	5s	476MB	⚠️ Unstable	
Component 1	✗	87	1s	516MB		
SLICE	✗	83	16s	713MB		
EiPiGraph linear	✗	81	1m	573MB		
PhenoPath	✗	79	5m	837MB		
pCreode	✗	78	2m	444MB	⚠️ Unstable	
Monocle ICA	✗	78	1m	692MB	⚠️ Unstable	
Wanderlust	✗	78	51s	413MB		
MATCHER	✗	77	43s	385MB		
Wishbone	✗	76	1m	370MB		
EiPiGraph cycle	✗	76	1m	532MB	⚠️ Unstable	
cellTree maptix	✗	74	4m	692MB	⚠️ Unstable	
MFA	✗	72	7m	669MB		
SCUBA	✗	70	5m	418MB	⚠️ Unstable	

Analysis workflow



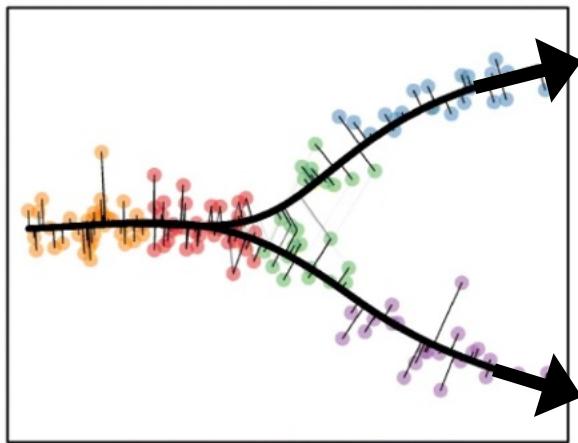
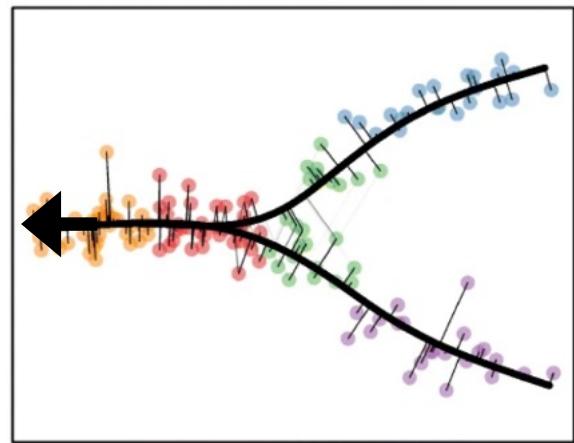
In Slingshot, the pseudotimes values are estimated as the euclidean distance from each point to the closest principal curve (i.e. by orthogonal projection of each point onto the curve).



But how are trajectories oriented???

A trajectory is not a vector: a trajectory is not oriented!

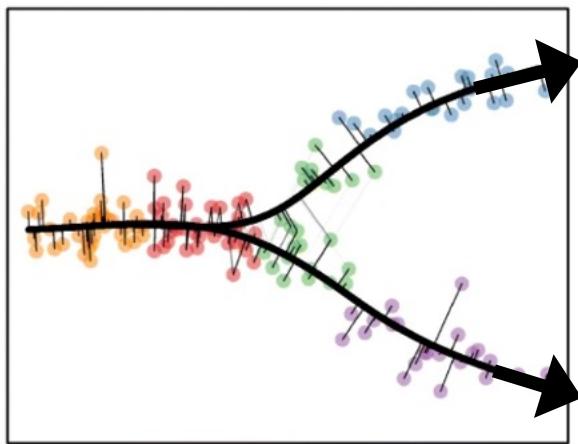
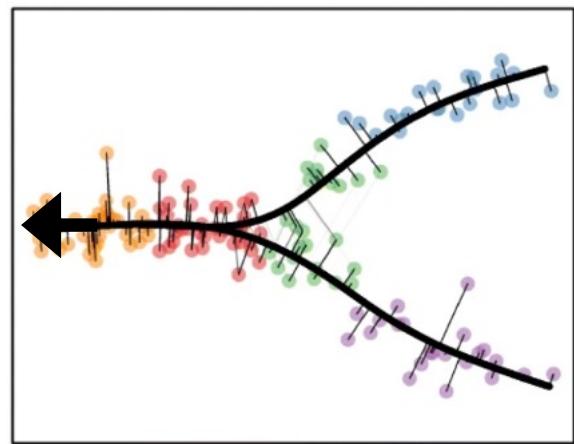
This is crucial when estimating pseudotime.



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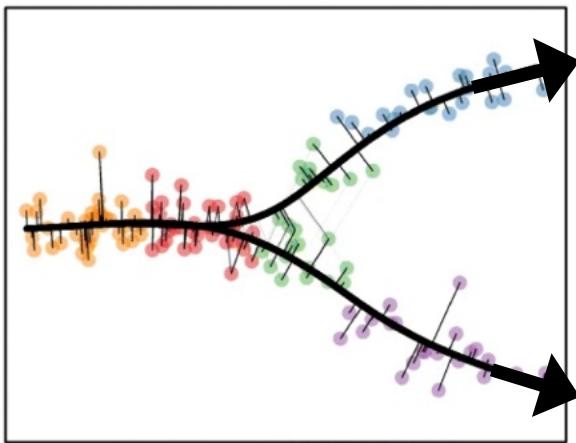
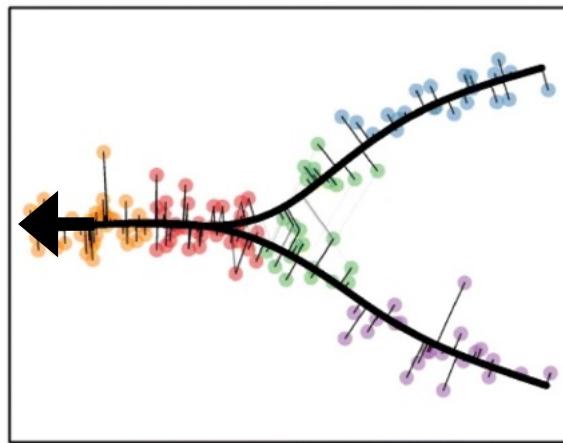


That is when one should use prior knowledge, ground-truth, or simply make a call.

But how are trajectories oriented???

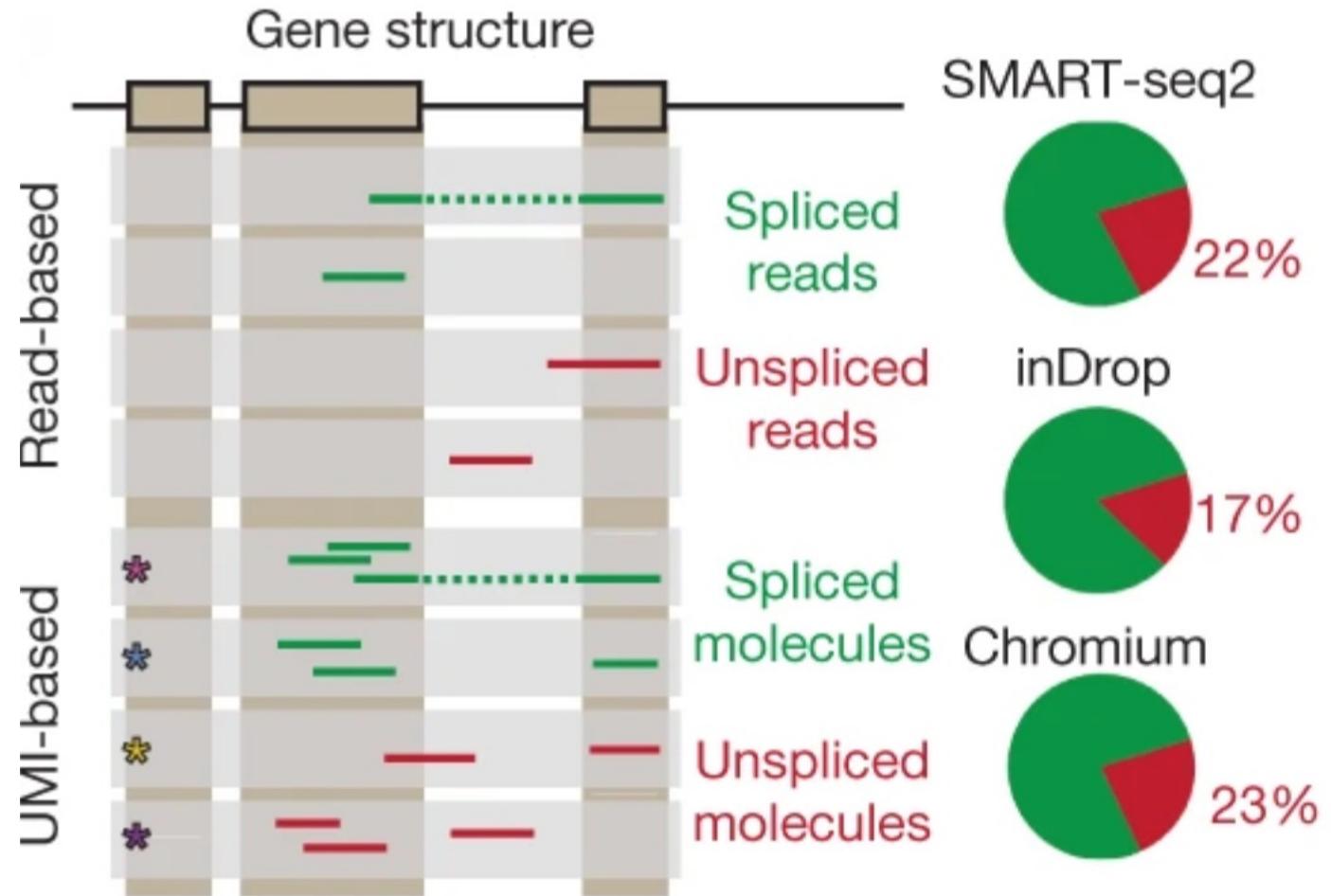
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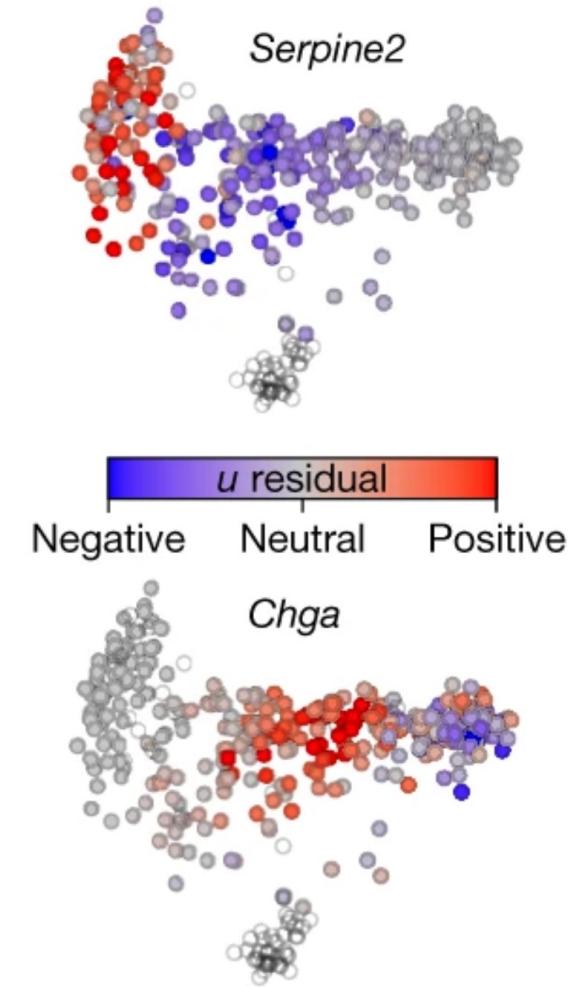
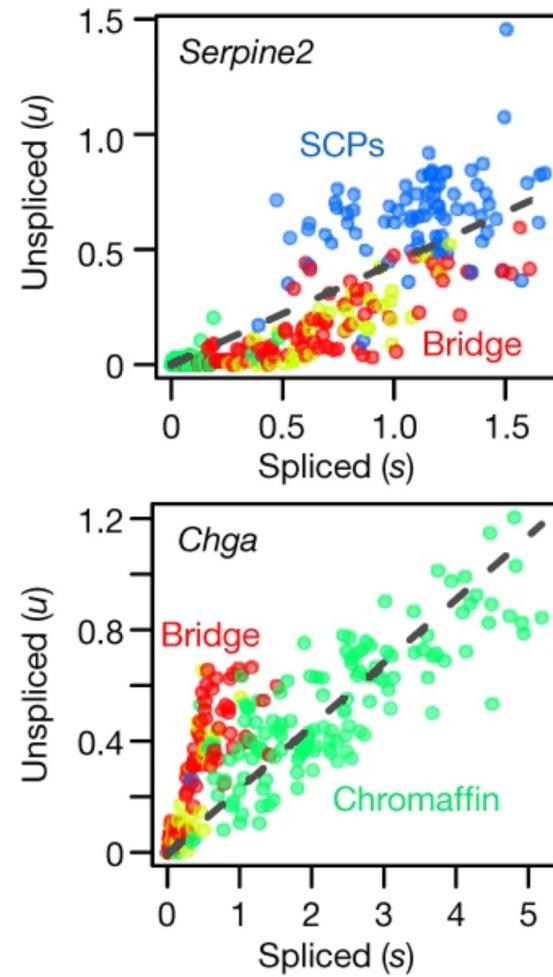
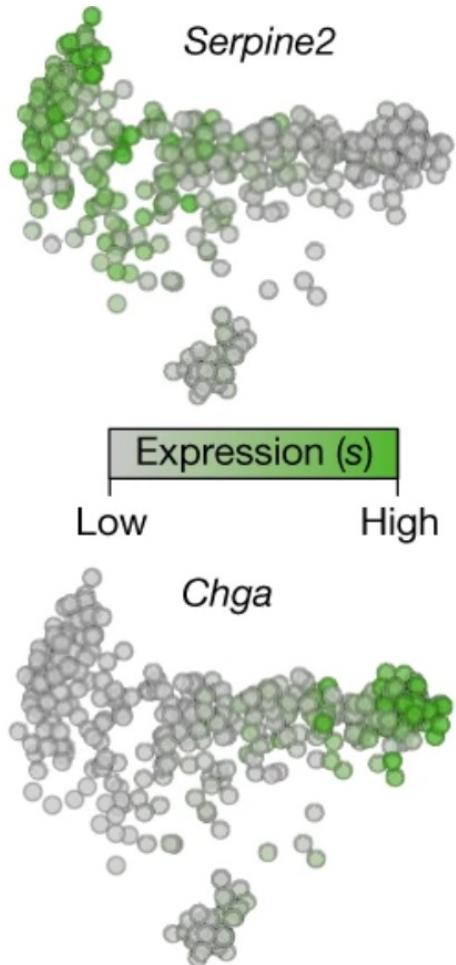
This is crucial when estimating pseudotime.



- Hematopoietic stem cell
- Lymphoid stem cell
- Committed lymphoid stem cell
- B cell
- T cell

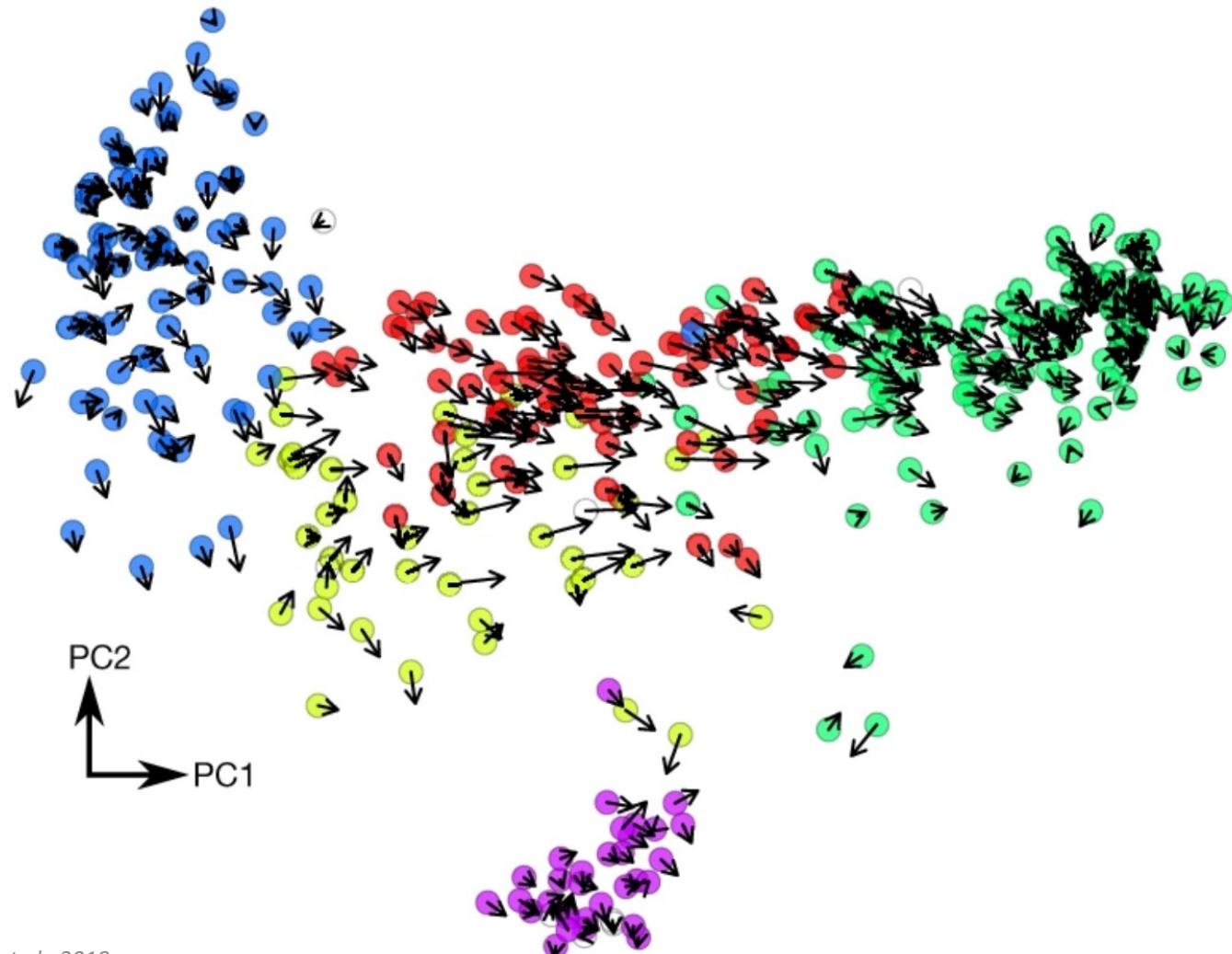
- Single-cell RNA-seq reads can be mapped onto exons or introns.



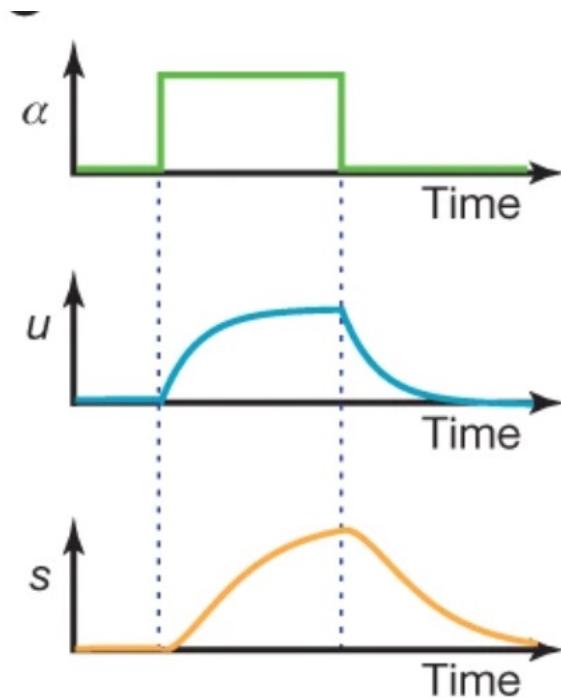
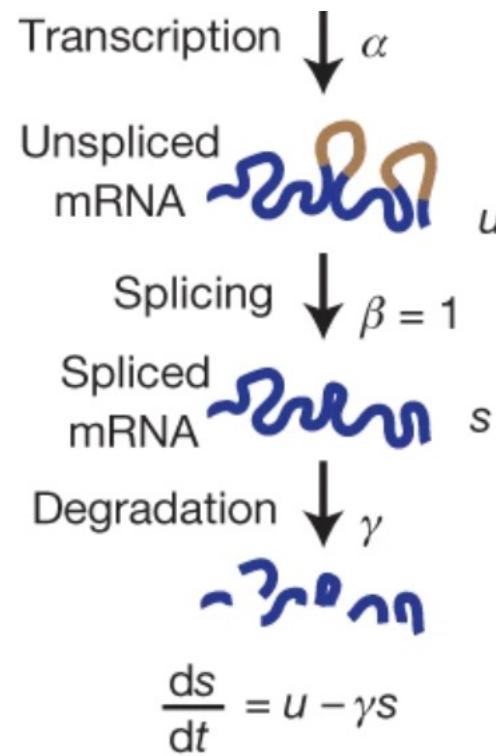


u residual represents whether the cell is far from an equilibrium state of expression

Generalizing this to all the detected (variable) genes, one can infer the future "position" of each cell.



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CAREFUL!!!

This model relies on an important assumption:

that transcription is steady-state.

This is rarely true, even less true for differentiating cells!!

Article | Published: 03 August 2020

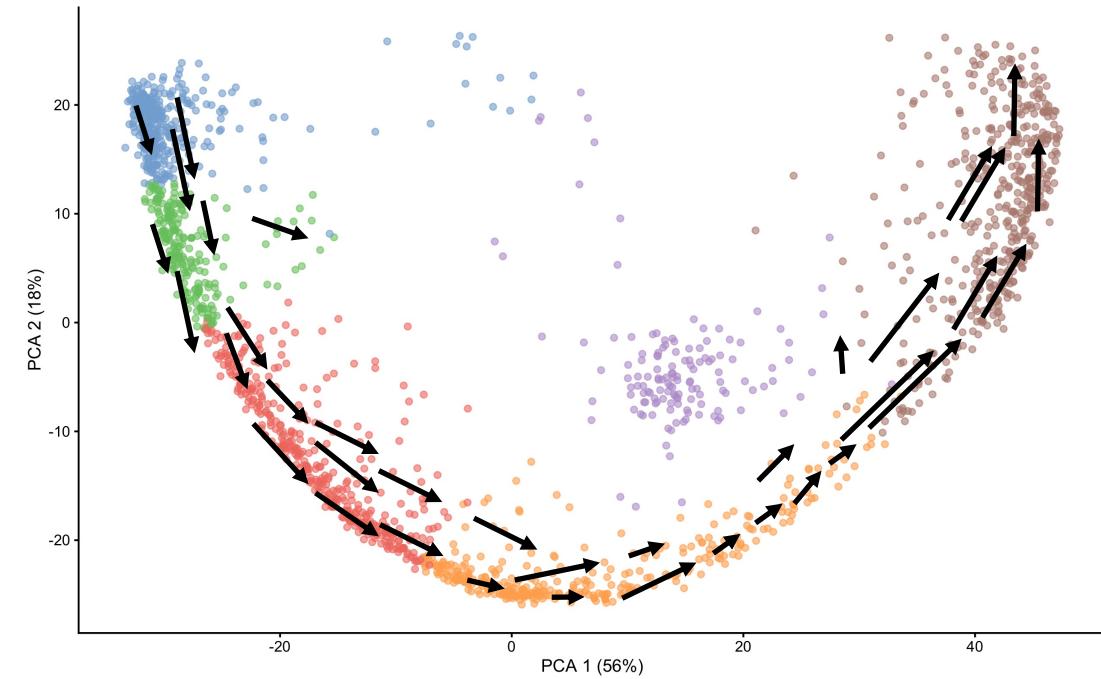
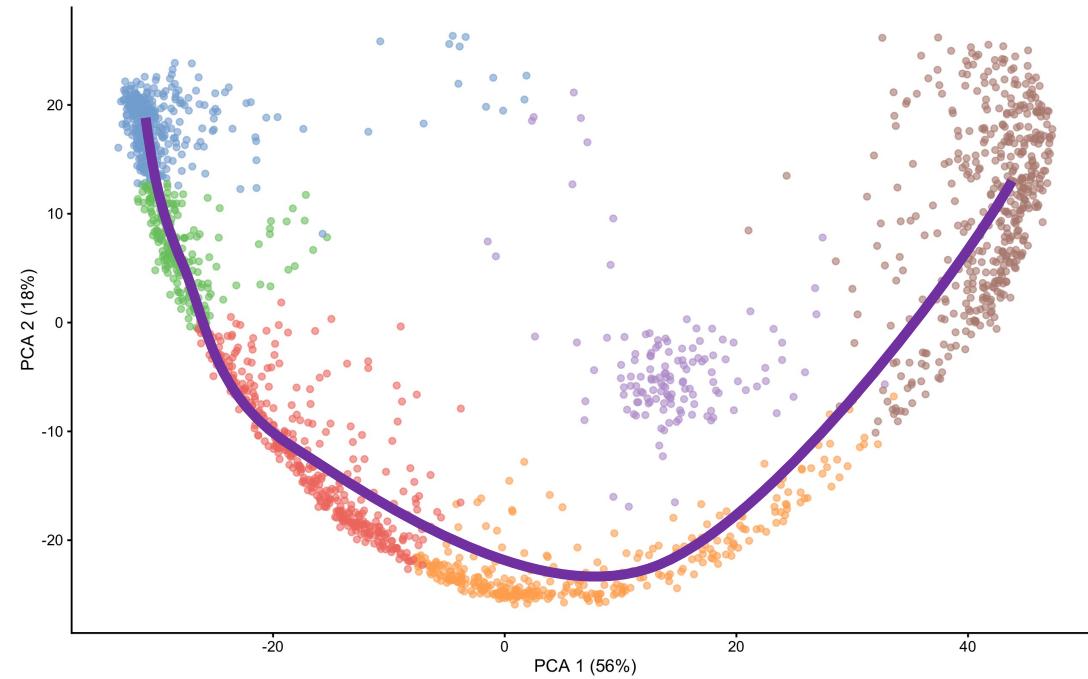
Generalizing RNA velocity to transient cell states through dynamical modeling

Volker Bergen, Marius Lange, Stefan Peidli, F. Alexander Wolf  & Fabian J. Theis 

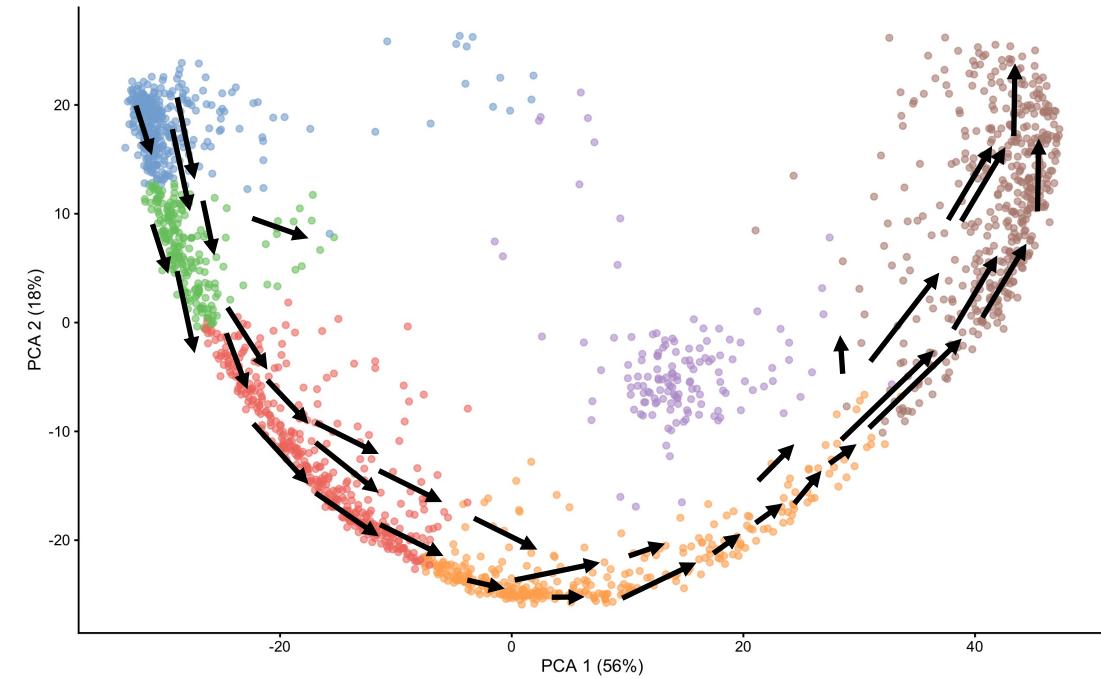
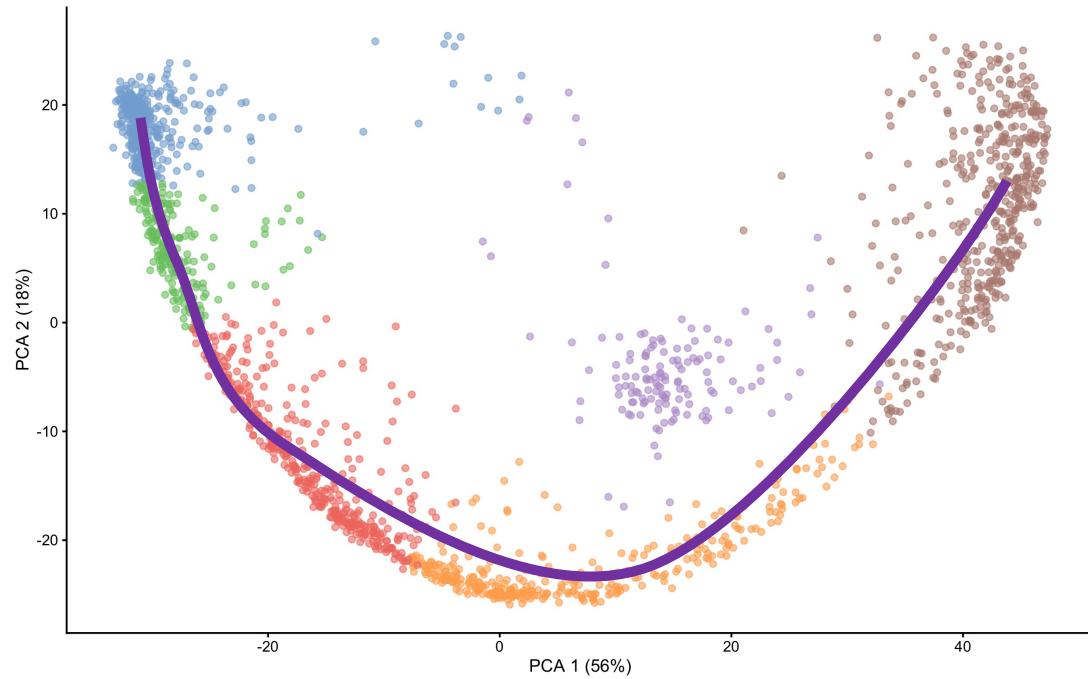
Nature Biotechnology **38**, 1408–1414 (2020) | [Cite this article](#)

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Using RNA velocity to infer directionality of the trajectory

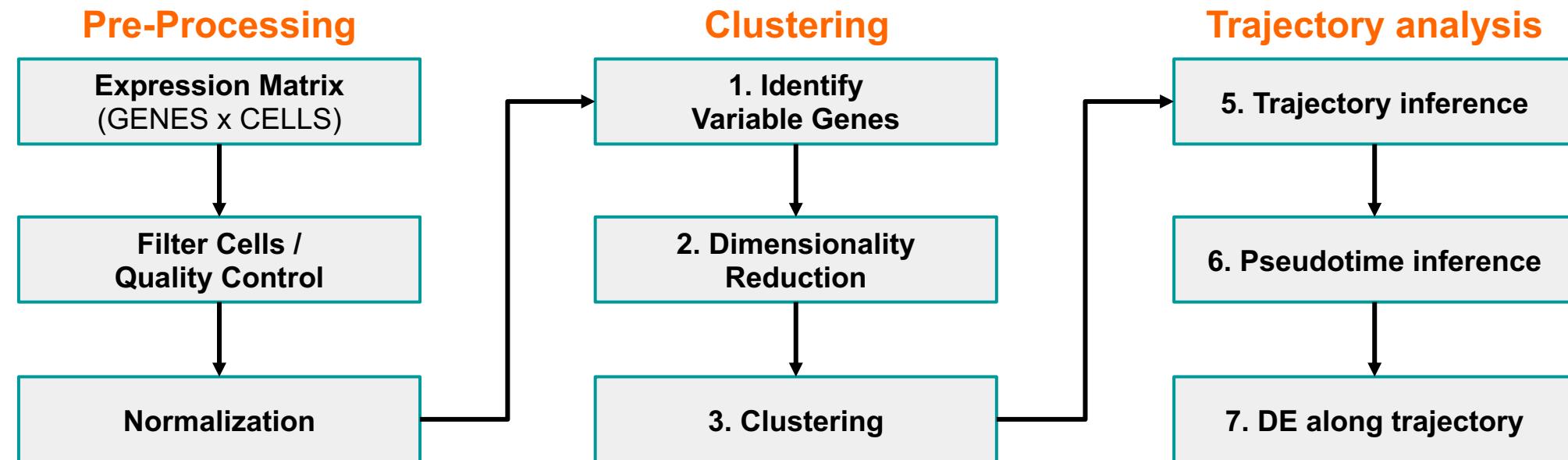


Using RNA velocity to infer directionality of the trajectory



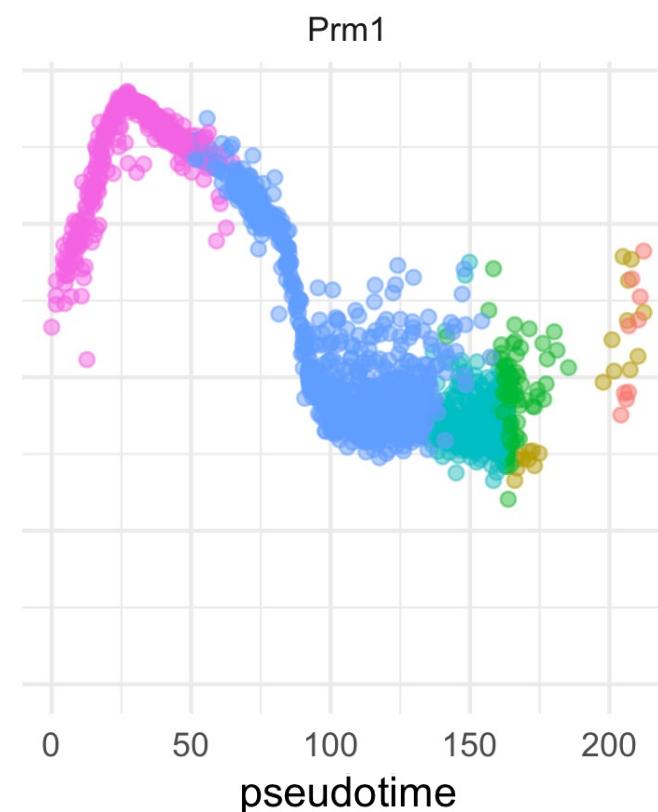
RNA velocity implementations generally output a pseudotime value. However, they do not infer trajectories!! So they cannot capture a branching event, or cycles, etc....

Analysis workflow



Modeling gene expression along pseudotime

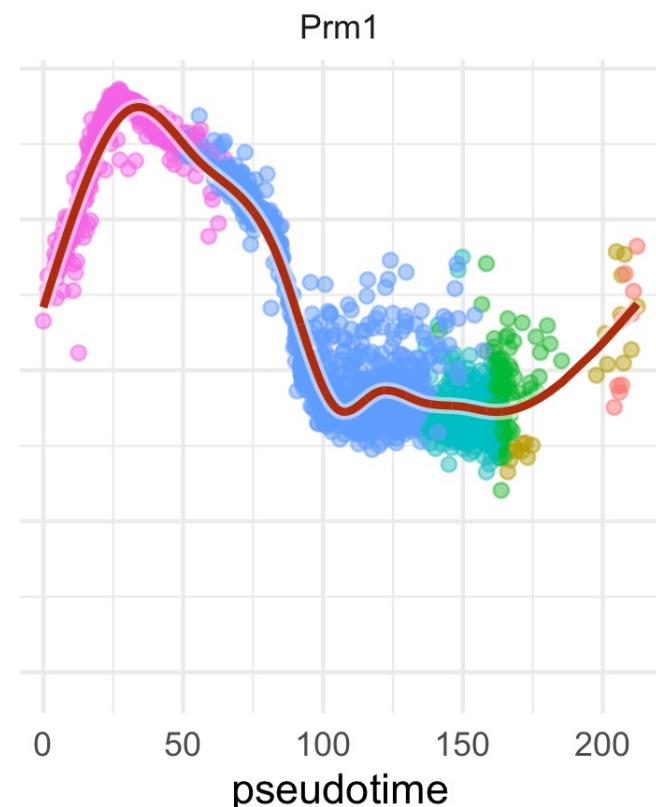
With a pseudotime value inferred to each cell, one can place all cells along an x axis, and plot gene expression on an y axis.



Modeling gene expression along pseudotime

With a pseudotime value inferred to each cell, one can place all cells along an x axis, and plot gene expression on an y axis.

Then time-dependent expression can be modelled, typically by fitting a GAM to the gene expression ~ pseudotime.



Performing temporal DE gene analysis

tradeSeq is an R package which facilitates GAM-fitting and gene DE analysis along and between trajectories.

It implements plug-and-play methods to use outputs from Slingshot, but most functions can be used directly with pseudotime values, without having to rely on trajectories specifically inferred with Slingshot.

Article | Open Access | Published: 05 March 2020

Trajectory-based differential expression analysis for single-cell sequencing data

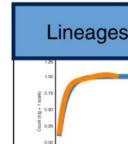
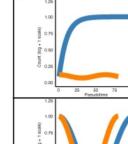
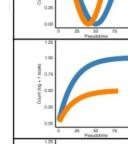
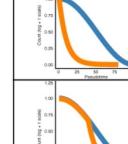
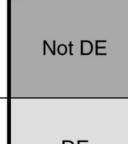
Koen Van den Berge, Hector Roux de Bézieux, Kelly Street, Wouter Saelens, Robrecht Cannoodt, Yvan Saeys, Sandrine Dudoit & Lieven Clement

Nature Communications 11, Article number: 1201 (2020) | [Cite this article](#)

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Van den Berge et al., 2020

c

Lineages	Within the orange lineage		Between the orange and blue lineages		
	association test	startVsEnd test	diffEnd test	pattern test	earlyDE test
	DE	DE	Not DE	Not DE	Not DE
	Not DE	Not DE	DE	DE	DE
	DE	Not DE	Not DE	Not DE	Not DE
	DE	DE	DE	DE	Not DE
	DE	DE	Not DE	DE	DE
	DE	DE	Not DE	DE	Not DE