

# ATAC-seq analysis

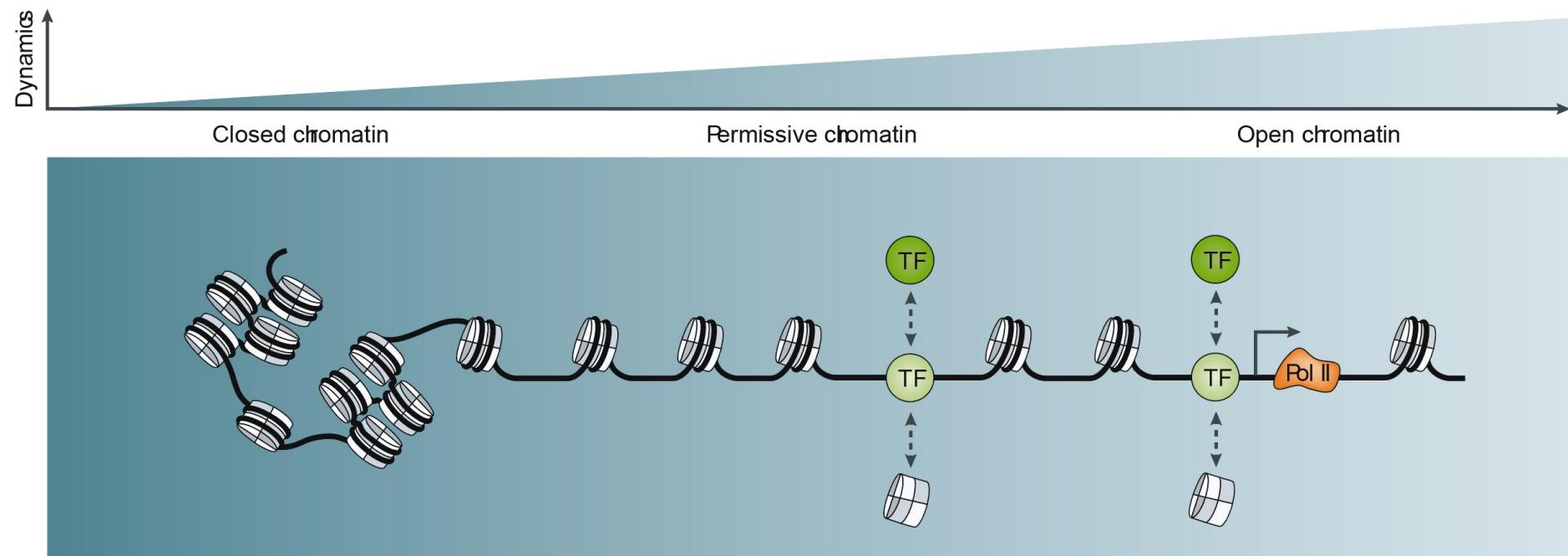
Epigenomics Data Analysis  
Jacques Serizay  
Physalia 2025



# Beyond textbook statements: a spectrum of regulatory capacity

Chromatin accessibility continuum that ranges from closed chromatin to highly dynamic, accessible or permissive chromatin

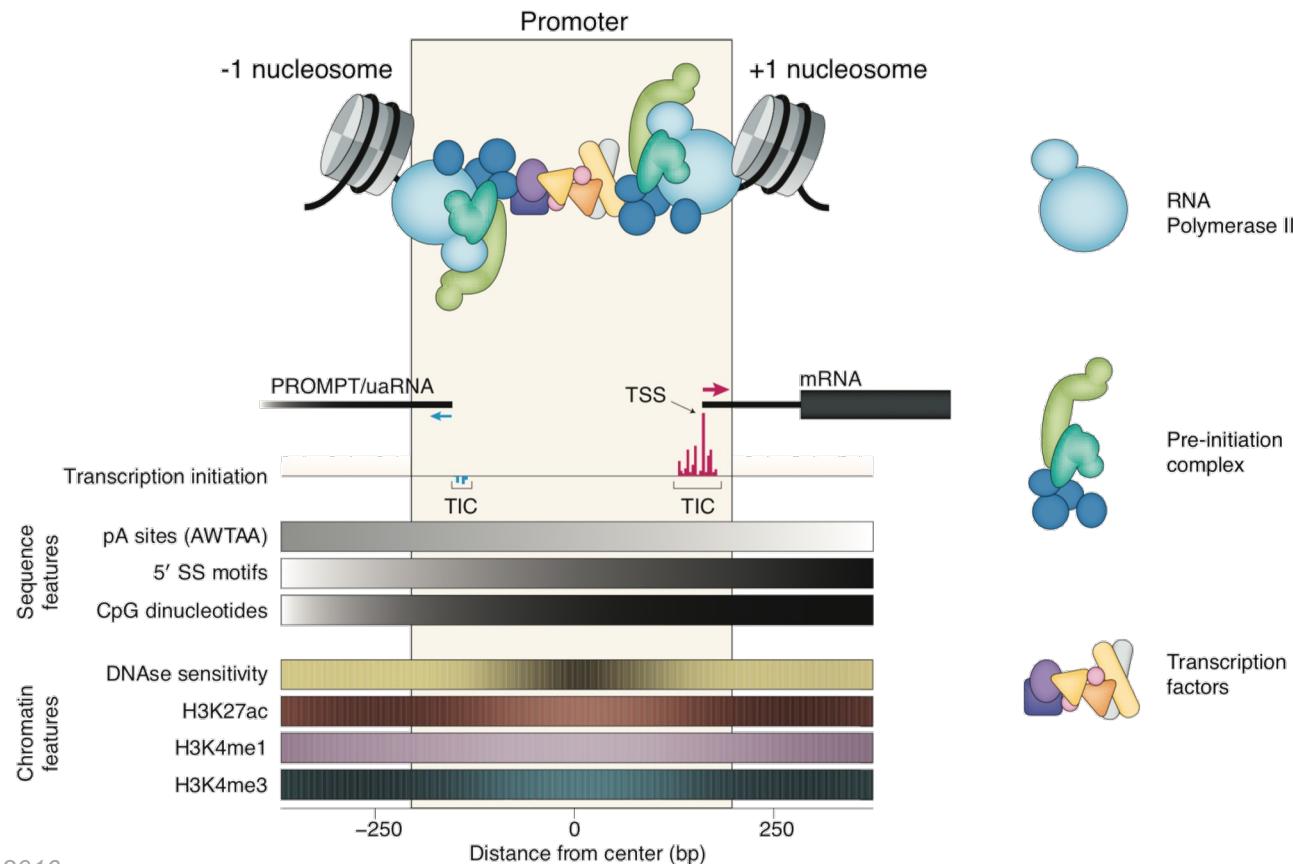
This landscape of chromatin accessibility reflects the spectrum of regulatory capacity — rather than a bistate organization



Klemm et al., Nat .Rev. Genet 2019

# Promoter organization

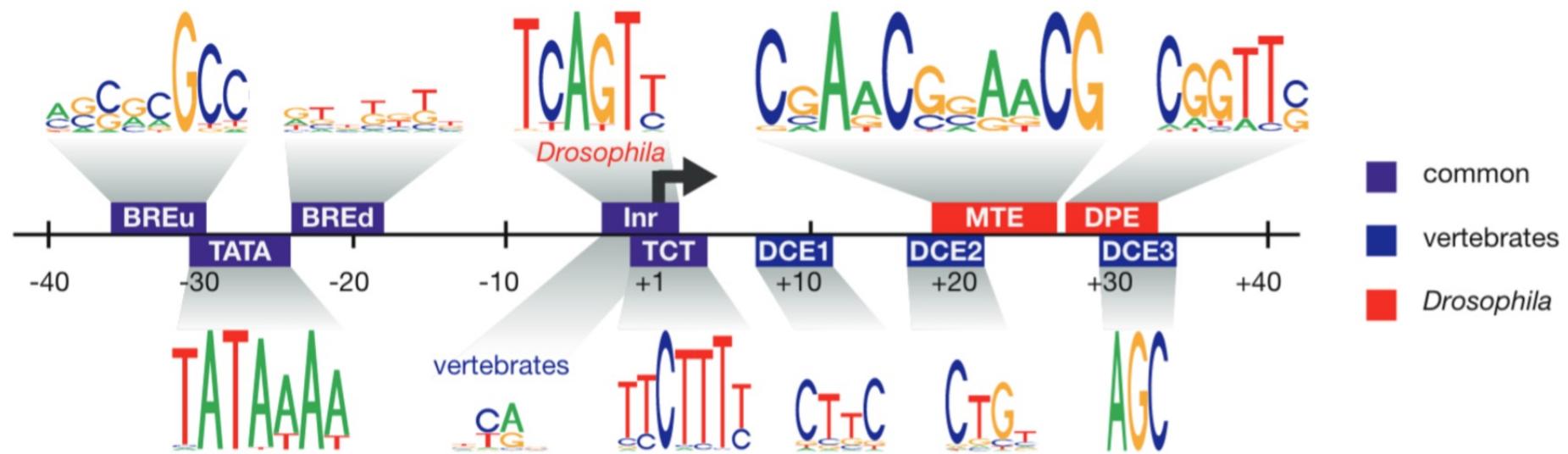
Promoters are crowded environments!



Haberle and Lenhard, *Semin. Cell Dev. Biol.* 2016

# Promoter organization

Transcription machinery and general transcription factors need access to DNA to recognize their binding motif

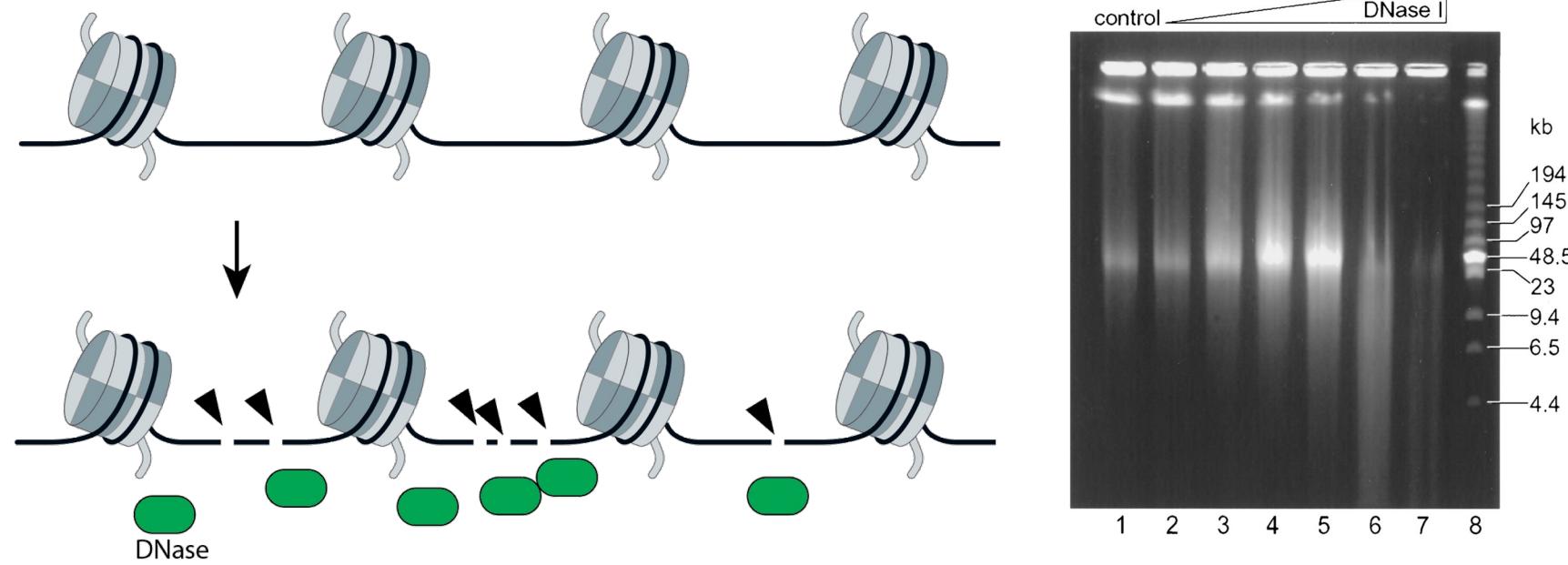


Haberle and Lenhard, Semin. Cell Dev. Biol. 2016

# How to measure chromatin accessibility: originally with nucleases

Nuclease enzymes were historically used to profile chromatin accessibility

- Dnase I (deoxyribonuclease I): a non-specific double-strand endonuclease

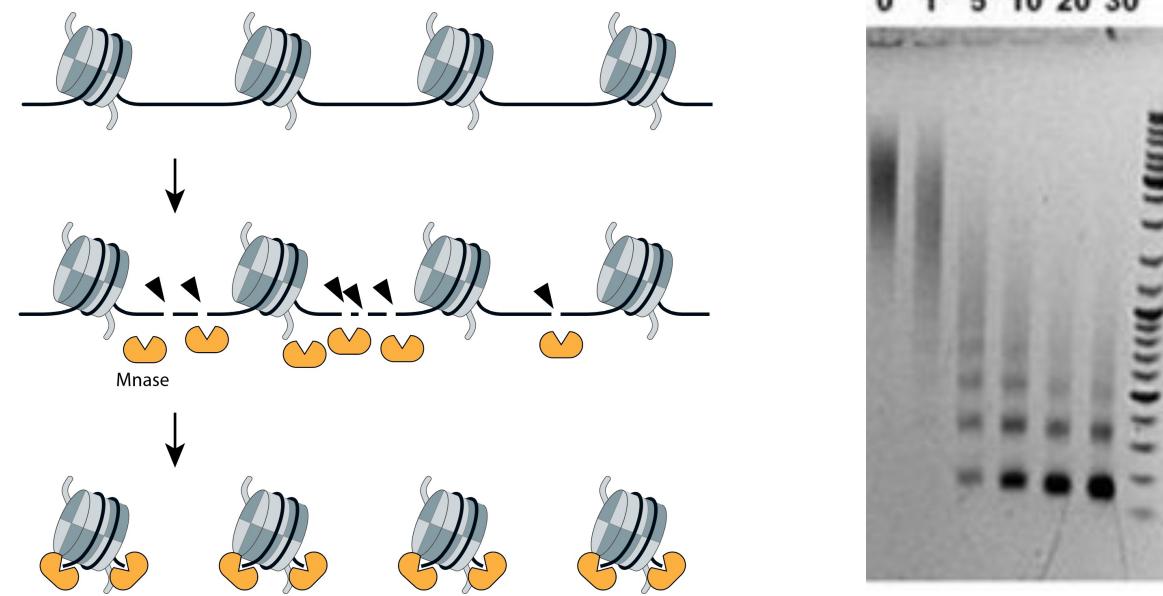


Paul & Ferl, *Plant Cell* 1998

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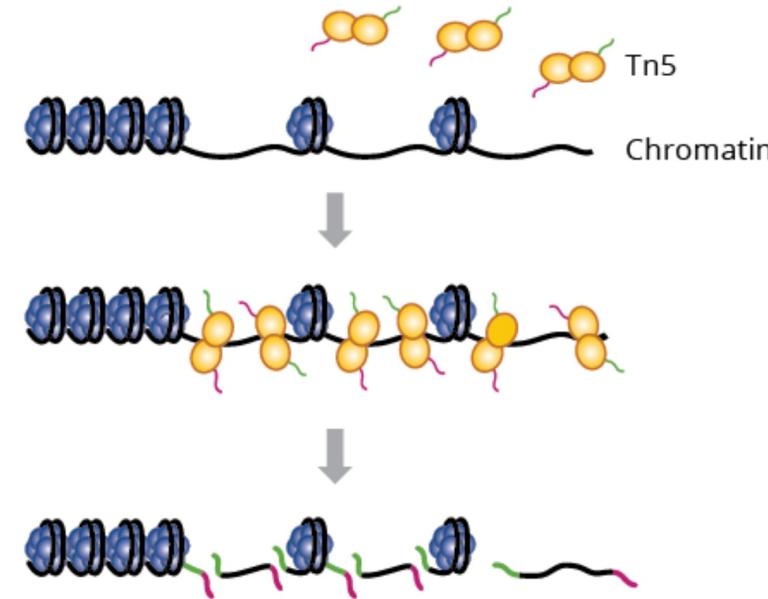


Rodríguez-Campos & Azorín, PLoS One 2007

# How to measure chromatin accessibility: originally with nucleases

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- Dnase I (deoxyribonuclease I): a non-specific double-strand endonuclease
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- ATAC: a Tn5 transposase, integrating transposons wherever it is possible (i.e. accessible)

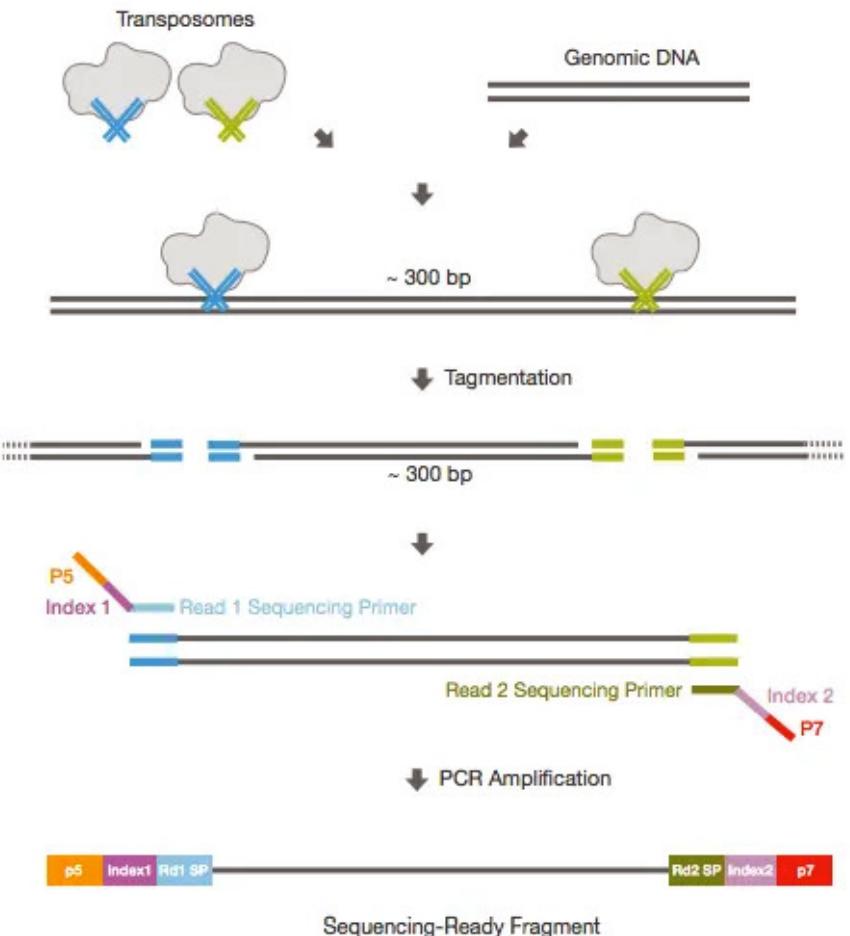


Buenrostro et al., 2013

# ATAC-seq: from chromatin to NGS library

Since the sequence of the transposons loaded on the Tn5 transposome is known, one can use them to in a PCR

- “Tagmented” (i.e. DNA with inserted transposons) will be amplified.
- Each end of a fragment corresponds to a transposition event.

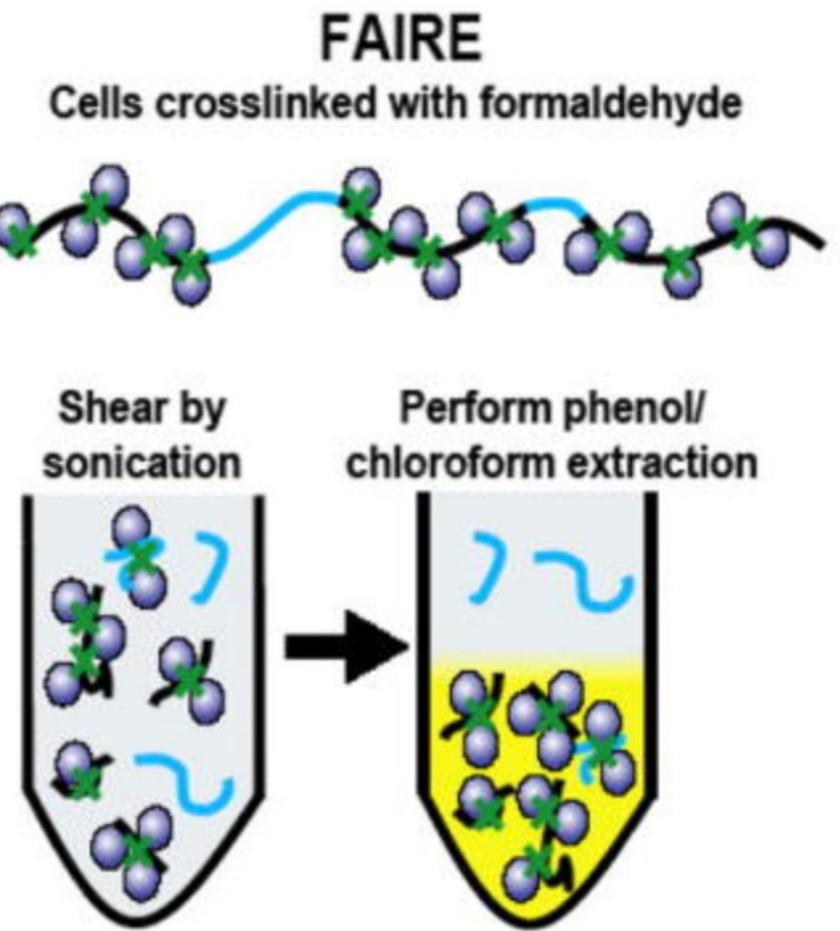


## Emergence of other enzymatic or mechanical approaches

### Mechanical approaches

- **FAIRE-seq**: Formaldehyde-Assisted Isolation of Regulatory Elements

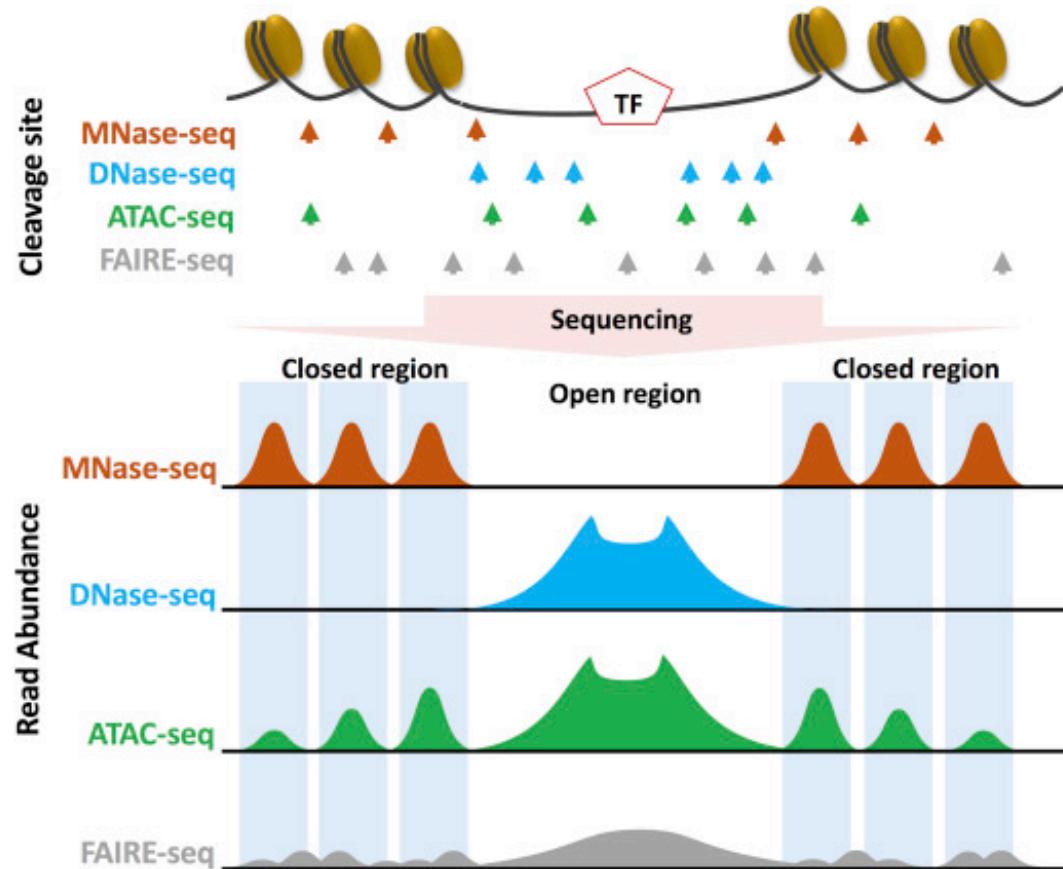
→ Uses a crosslinking + sonication + phenol extraction to isolate nucleosome-depleted chromatin



Giresi & Lieb, Methods 2009

# Comparison of the main experimental approaches

Each assay generates a specific type of profile.

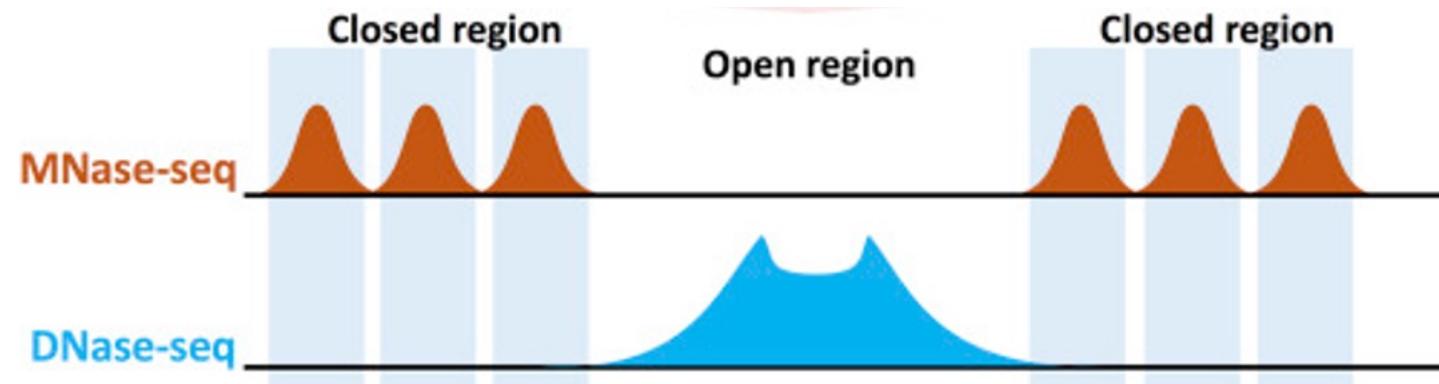


Hsu et al., Epigenetics in Human Diseases 2018

## Positive vs. negative measurements

DNase-seq relies on presence of signal (**positive measurements**) to map accessible regulatory elements

Mnase-seq relies on absence of signal (**negative measurements**) to map accessible regulatory elements



# ATAC-seq downstream analysis



Get .bcl files



Create fastq files



Or **bcl2fastq**



QC: remove/trim low quality reads

E.g. **cutadapt**



Align fastq to BAM

E.g. **bowtie2**



Filter duplicates, artifacts, ...

E.g. **samtools**



Generate tracks

E.g. **deepTools**



Assay-specific downstream analysis

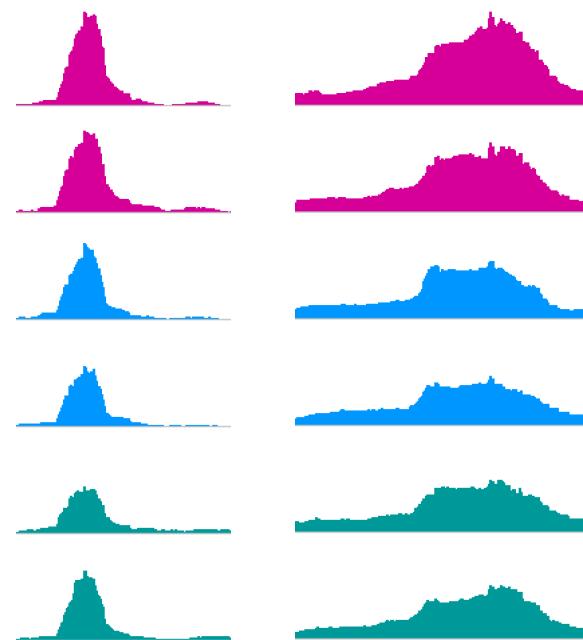
**Peak calling**  
**Differential peak analysis**  
**Fragment size distribution**  
**Promoter footprint analysis**

## ATAC peak callers

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Sample 1

Sample 2

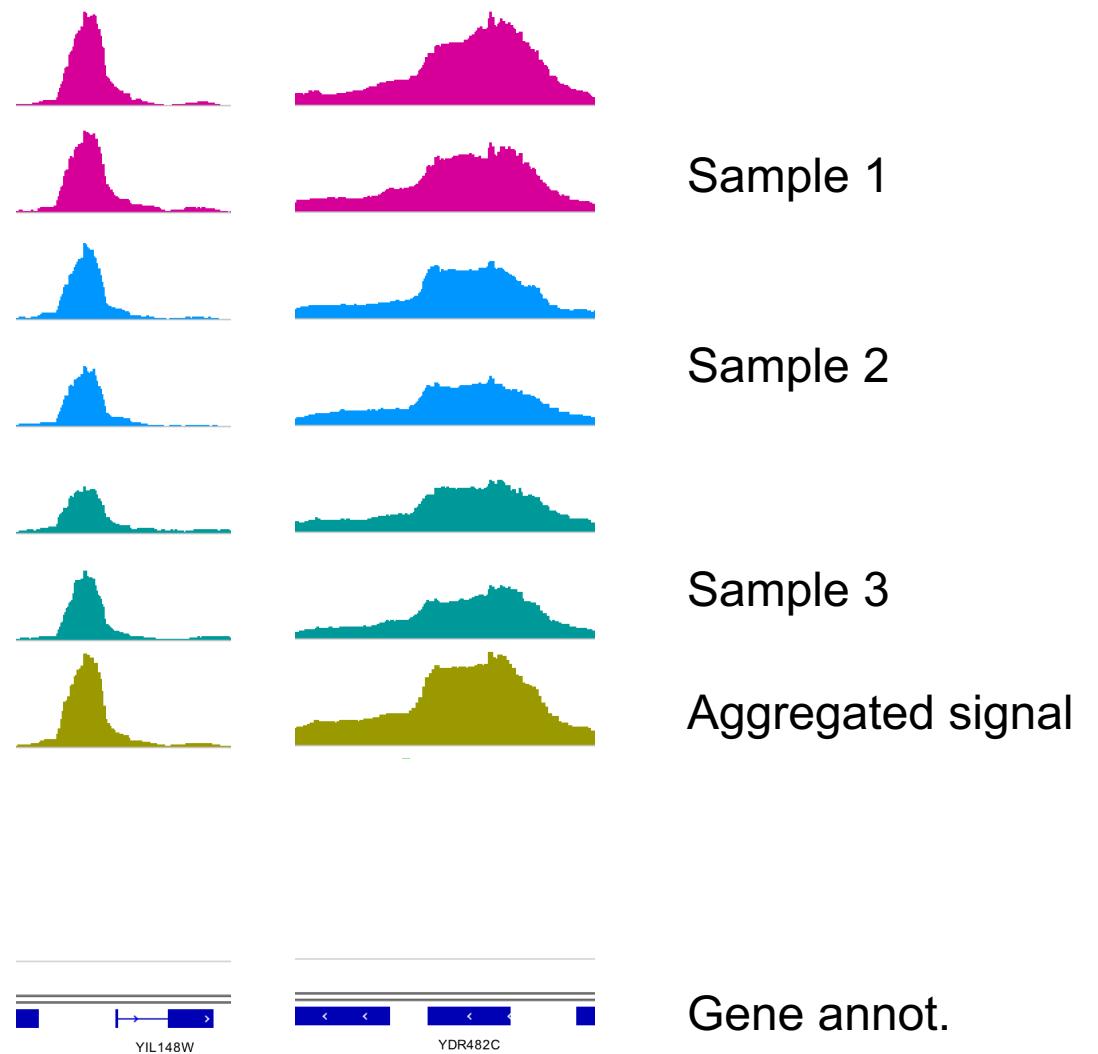
Sample 3



Gene annot.

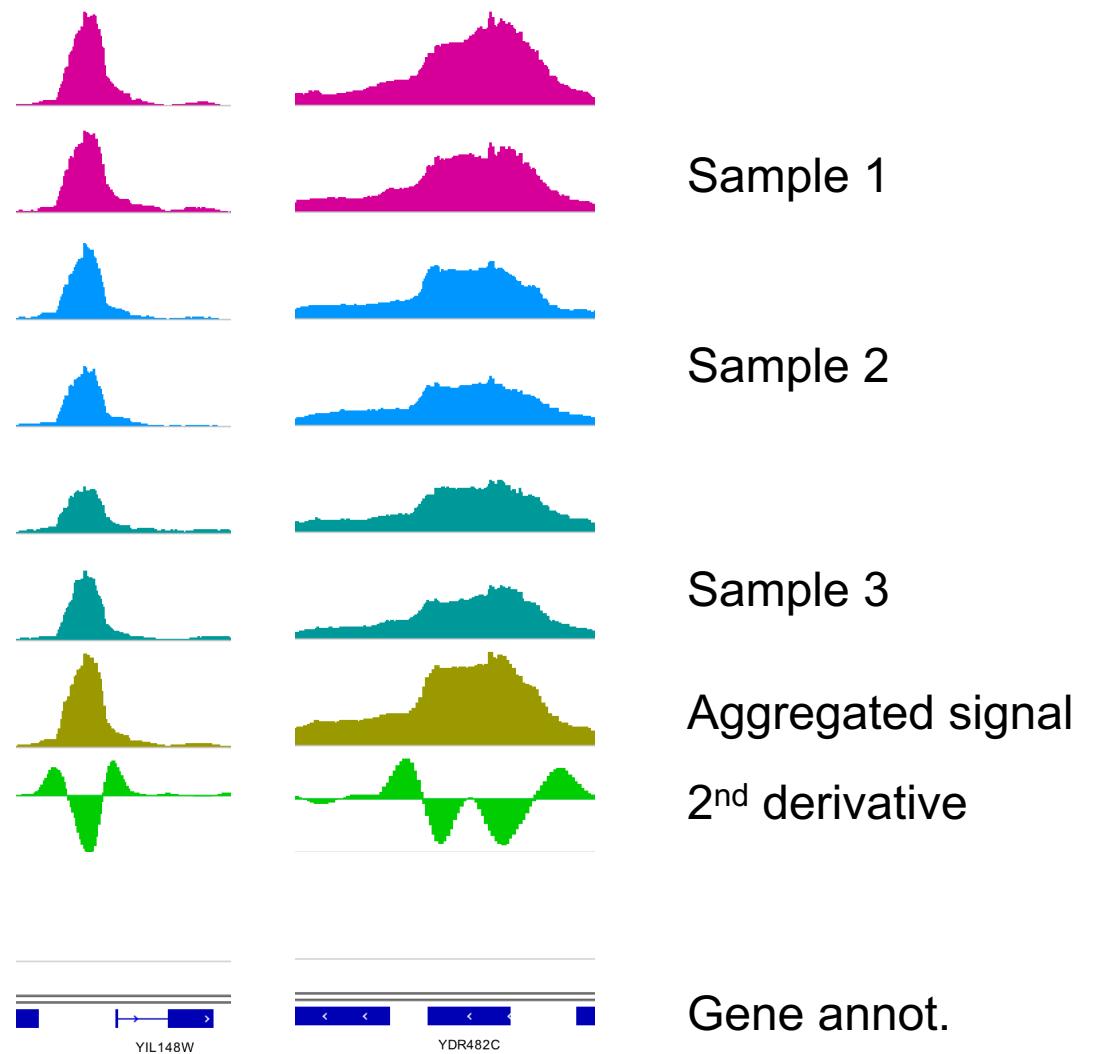
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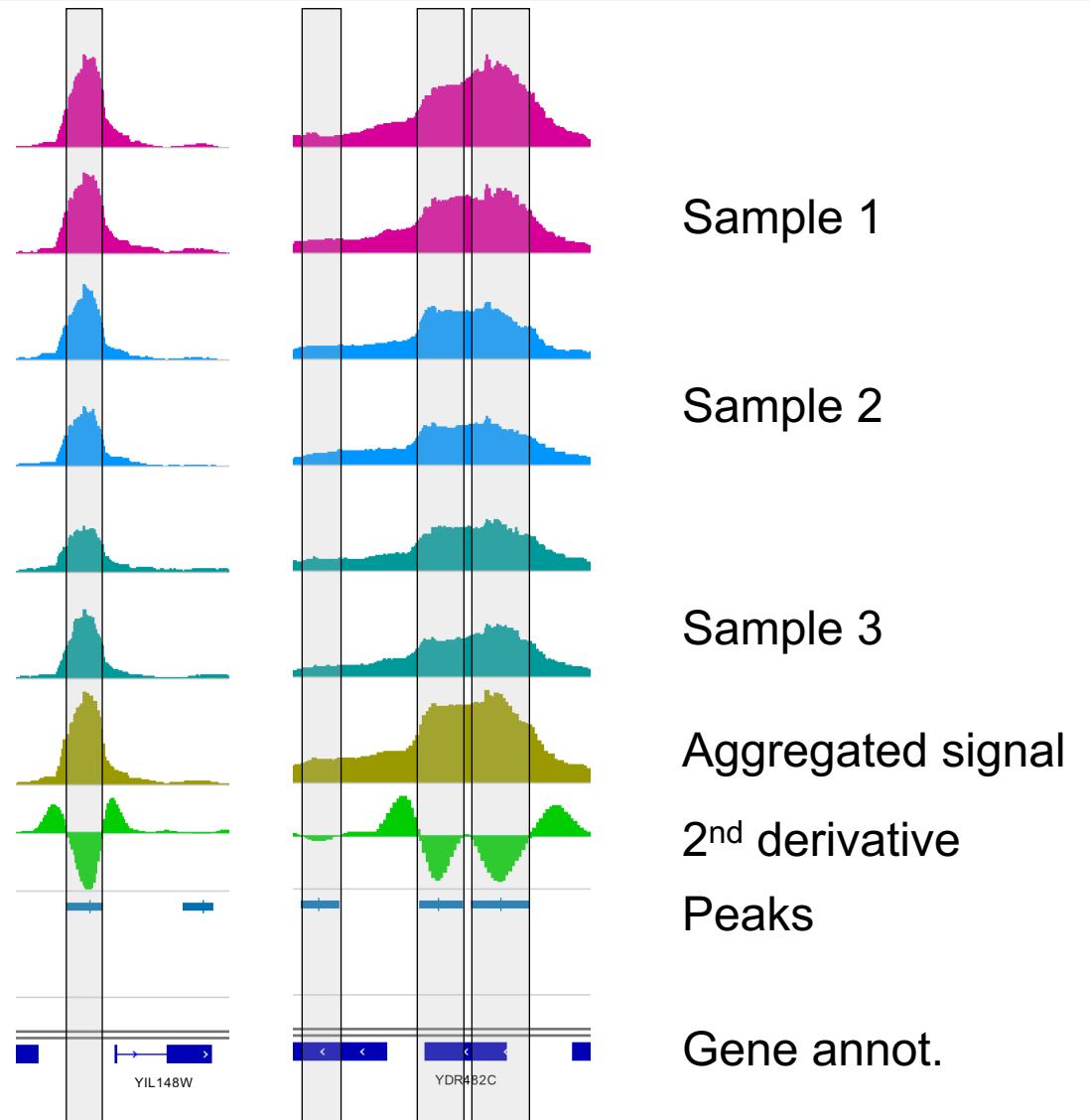
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```
yapc output_refix \
    sample1 sample1_rep1.bw sample1_rep2.bw \
    sample2 sample2_rep1.bw sample2_rep2.bw \
    ...
    sampleN sampleN_rep1.bw sampleN_rep2.bw
```